



WORKSHOP
CLIMATE CHANGE AND SEEDS
SEEDS FOR THE FUTURE
Chania (Greece), Sep. 25-26, 2007



Climate Change and Seed Germination - a scientific and technical approach

Costas Thanos - UNKA



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SEEDS FOR THE FUTURE
Chania (Greece), Sep. 25-26, 2007



SEED: the most **tolerant**
stage in the life cycle of
a plant

SEEDLING: the most
vulnerable stage in the
life cycle of a plant



Dormancy types vs Germination strategies

Instead of studying Dormancy Types we should concentrate on Germination Strategies



The GERMINATION STRATEGY

is a long term plan of action designed to achieve the particular goal of the successful transition from the seed stage to the seedling stage.





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GERMINATION ADAPTIVE TRAITS I
Chania (Greece), Sep. 24-26, 2007




A GERMINATION STRATEGY encompasses various adaptations (or adaptive traits) at the structural and physiological level during:

seed maturation

post-dispersal / pre-germination

germination (s.s.)





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GERMINATION ADAPTIVE TRAITS I (MATURATION)

- Life history of seed maturation
- Conditions of seed maturation
- Desiccation tolerance





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GERMINATION ADAPTIVE TRAITS II

CHATEAU D'AVIGNON, SEP. 25-28, 2007




GERMINATION ADAPTIVE TRAITS II (POST MATURATION)

- Dispersal
- Predation
- Soil and canopy seed banks
- Seed viability





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GERMINATION ADAPTIVE TRAITS III

CHATEAU D'AVIGNON, SEP. 25-28, 2007




GERMINATION ADAPTIVE TRAITS III (PREREQUISITES)

- An undeveloped (or underdeveloped) embryo
- An 'after-ripening' requirement
- Hard (water-impermeable) seed/fruit coats
- Chemically inhibiting substances in the germination unit
- A (cold or warm) stratification need

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GERMINATION ADAPTIVE TRAITS IV
(GERMINATION s.s.)
Chambre MATISOL, Sep. 24-28, 2007

GERMINATION ADAPTIVE TRAITS IV (GERMINATION s.s.)

- Temperature range
- Light quality and quantity
- Speed (rate) of germination
- Osmotic and salinity stress
- Promoting or inhibiting chemicals
- Season of field germination

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 China (WATON), Sep. 24-28, 2007




6.5 Effects of climate change

The consequences of climate change for regeneration from seeds are likely to be complex and far-reaching, and some effects go beyond what can reasonably be considered here.

Regeneration from seed is generally expected to benefit from climate warming, but perhaps this is because attention has tended to focus on the colder parts of the globe. For example, alpine populations of *Gentianella germanica* may set hardly any seed in cool years and would be expected to produce more regular seed crops in a warmer climate (Wagner & Mitterhofer, 1998). Similarly, the phenology of *Dryas octopetala* was advanced by experimental warming at four tundra sites (Welker et al., 1997). Enhanced seed maturation, germination and seedling survival of the only two native Antarctic vascular plant species (*Colobanthus quitensis* and *Deschampsia antarctica*) seems to be a more sensitive indicator of climate change in west Antarctica than the response of the dominant cryptogams (Smith, 1994). However, improved seed production does not necessarily translate into larger populations of Arctic and alpine plants. Neither alpine *Ranunculus acris* nor three Arctic and alpine populations of *Saxifraga oppositifolia*





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Une traité récente synthétique du sujet (Fenner & Thompson 2005) a conclut que les conséquences du changement climatique sur la régénération par semences sont probablement très compliquées. Quelques aspects de la **biologie reproductive** que puissent être impliquées et influencées sont: l'augmentation ou la diminution de la **régularité de la production des semences** aux plantes alpines ou maritimes, respectivement, des types altérés de **'masting'** aux arbres et arbustes, l'augmentation ou la diminution de la **production des semences** aux limites extrêmes de la distribution d'un espèce avec une changement parallèle de la distribution, le **délai ou l'inhibition complète de la germination** et de **l'émergence de la plantule** aux conditions de plus en plus arides de la saison de la recrutement habituelle (automne ou printemps), l'échec (ou **timing inapproprié**) de recevoir des 'doses' suffisantes des exigences climatiques spécifiques, comme post-maturation, stratification etc que induisent la germination.





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Effects of climate change on:

- Flowering and reproduction (in general)
- Seedling survival and establishment
- Seed yield and maturation (incl. masting)
- Soil seed banks
- **Seed germination requirements**





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Effects of climate change on Seed germination requirements:

- Underdeveloped embryo
- Afterripening
- Hard seed coat
- Soil moisture (for recalcitrant seeds)
- Stratification
- Rate (speed) of germination
- Temperature

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[Cistus \(Fire-adapted Rockroses\)](#)
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 Chania (Greece), Sep. 24-28, 2007

CASE STUDY I

Cistus spp.
(fire-adapted rockroses)

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Semclimes

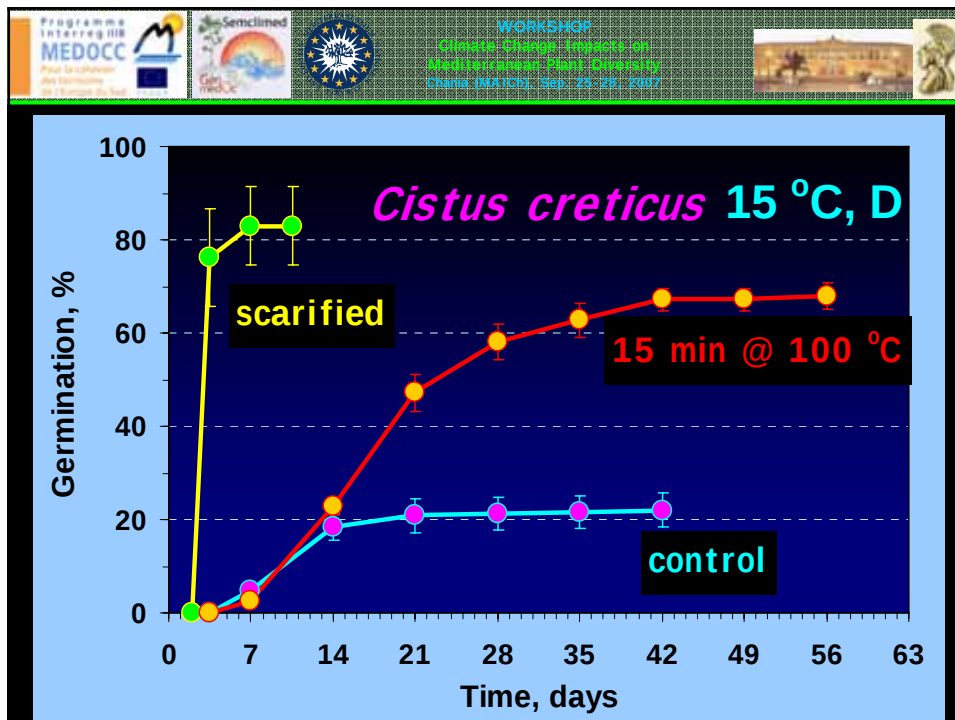
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[Cistus \(Fire-adapted Rockroses\)](#)
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Cistus creticus

Cistus parviflorus

Cistus salvifolius



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Cistus creticus: 1000 ans
de méditerranéisme
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Moreover, (in *Cistus creticus*) the promotive effect caused by fire or thermal shock seems to be clearly different from that brought about by mechanical scarification.

This latter treatment results in an impressive enhancement of germination which is completed in less than 1 week. On the other hand, a thermal pretreatment does allow the penetration of water but the eventual germination is markedly slow.

(Thanos & Georgiou, Plant, Cell & Environ. 1988)

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Chambre (MATHON) - Sep. 24-25 - 2007

CASE STUDY II

Coridothymus capitatus
(thyme)

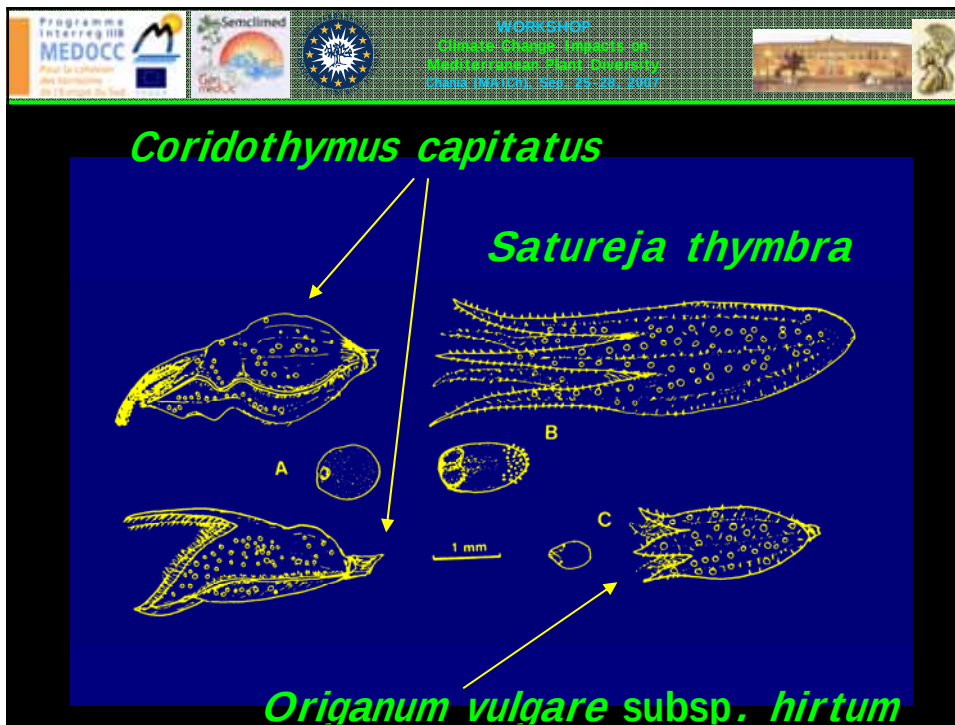
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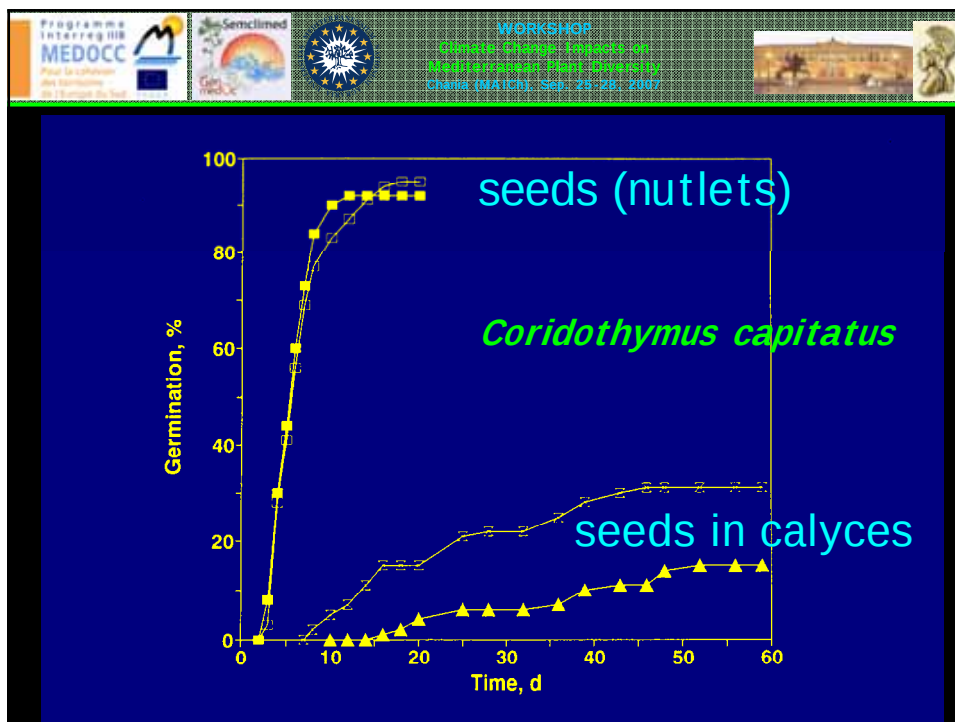
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Coridothymus capitatus





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CORRIDOR FOR LIFE: COOPERATION
BIOLOGICAL CORRIDOR FOR LIFE
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CASE STUDY III

Aesculus hippocastanum (the Balkan Horse-chestnut)

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Genetic diversity of *Aesculus hippocastanum*
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Aesculus hippocastanum




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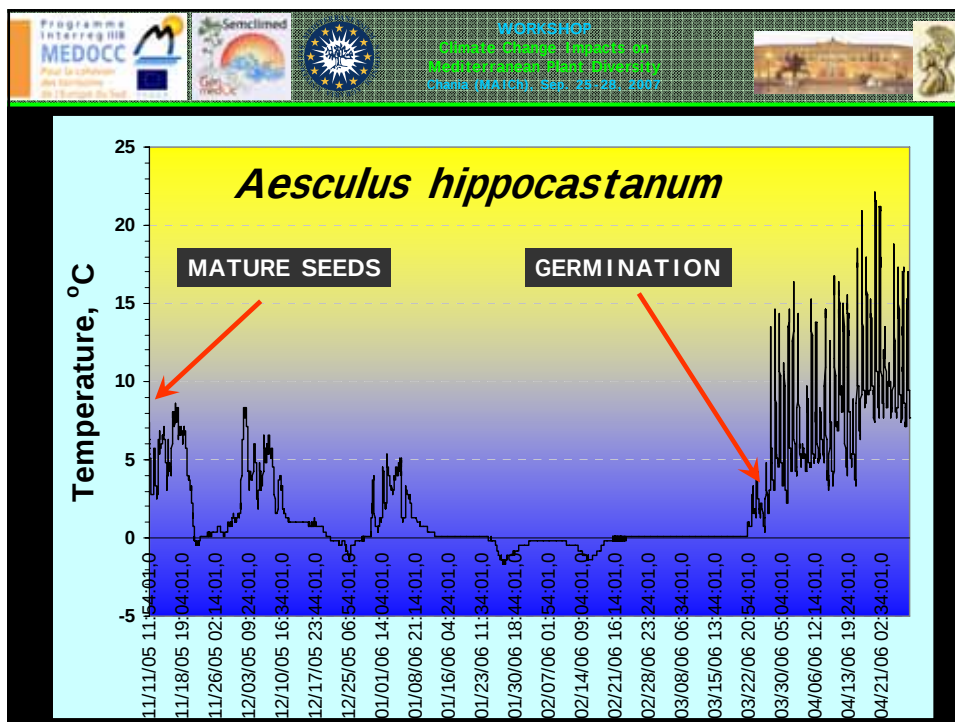
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Genetic diversity of *Aesculus hippocastanum*
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Aesculus hippocastanum

Water content of seeds: 52.3 % (fwb)

Seeds can germinate without any external water supply after a period of 6-month storage at 5 °C





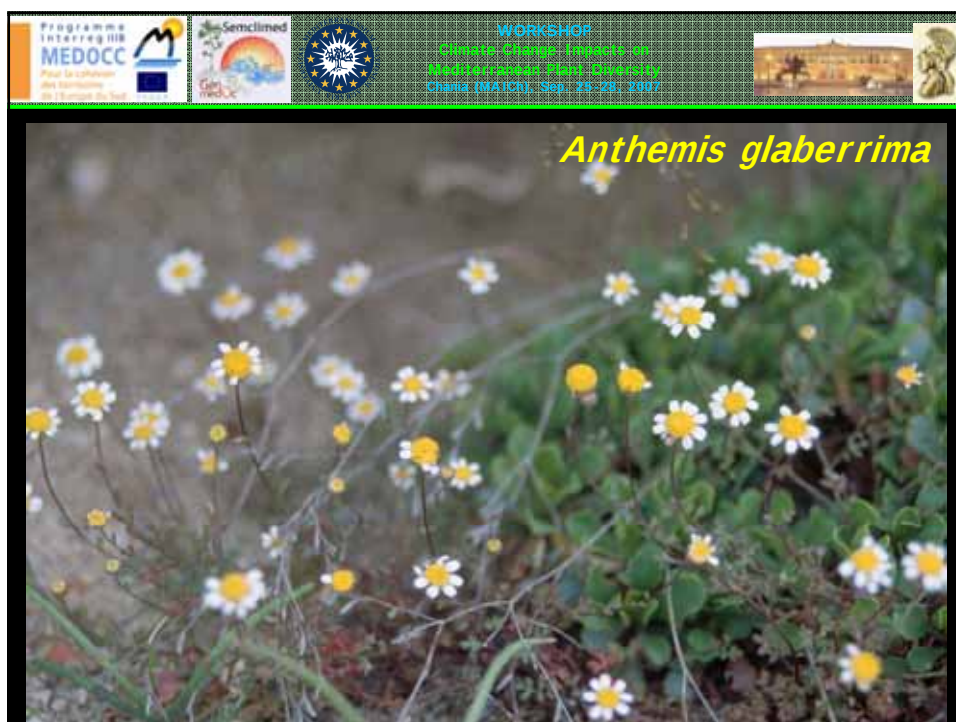
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MEDOCC 2007-2013
MEDOCC 2007-2013
OASIS (MATCH) Sep. 24-28, 2007

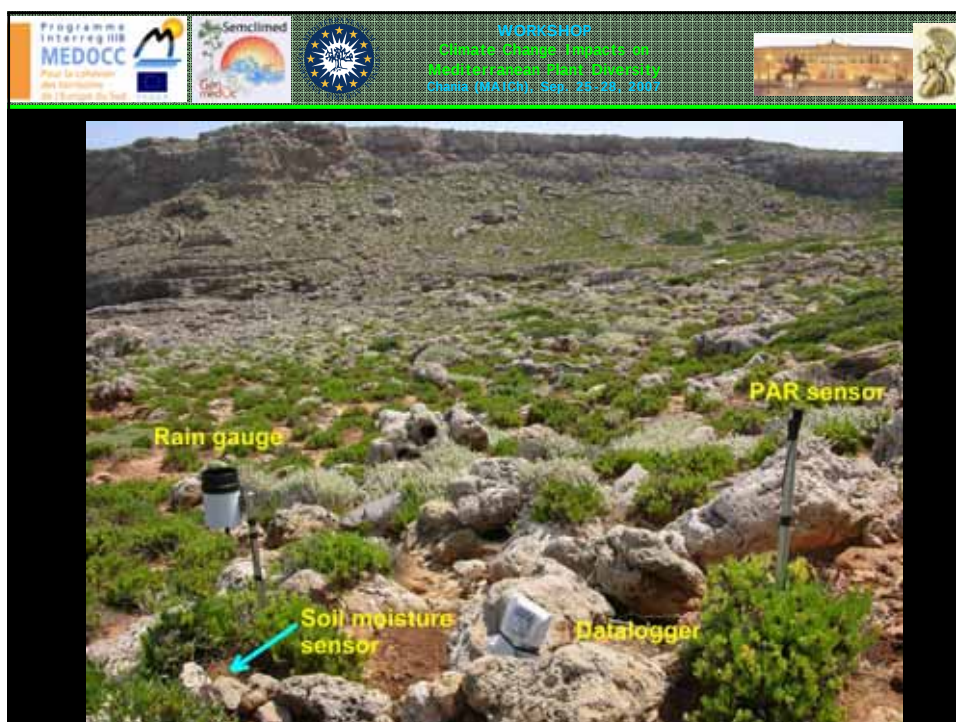
CASE STUDY IV

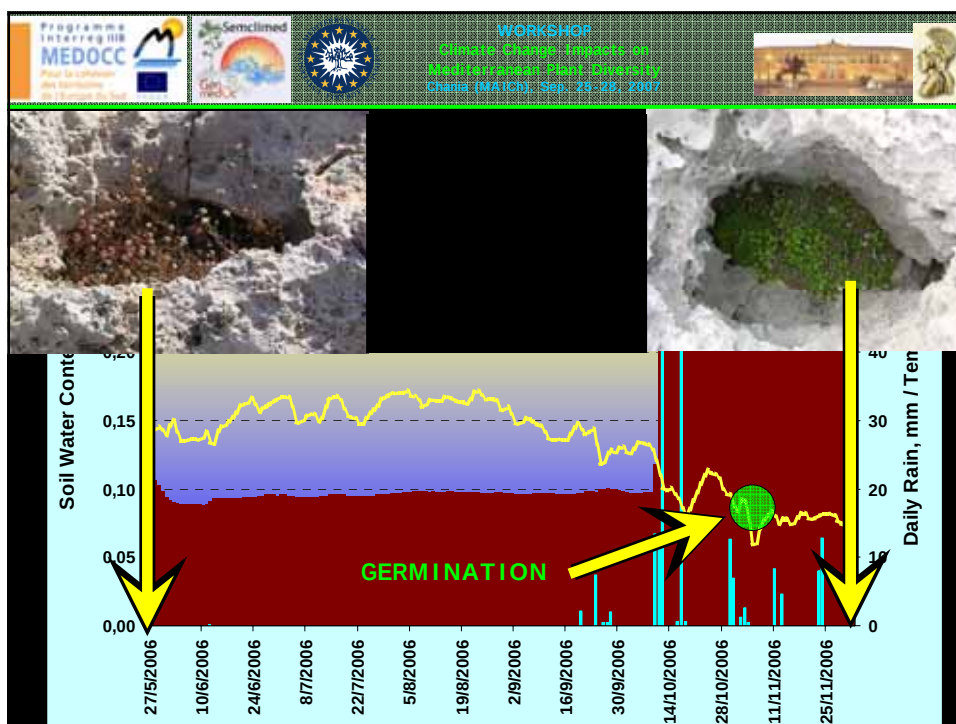
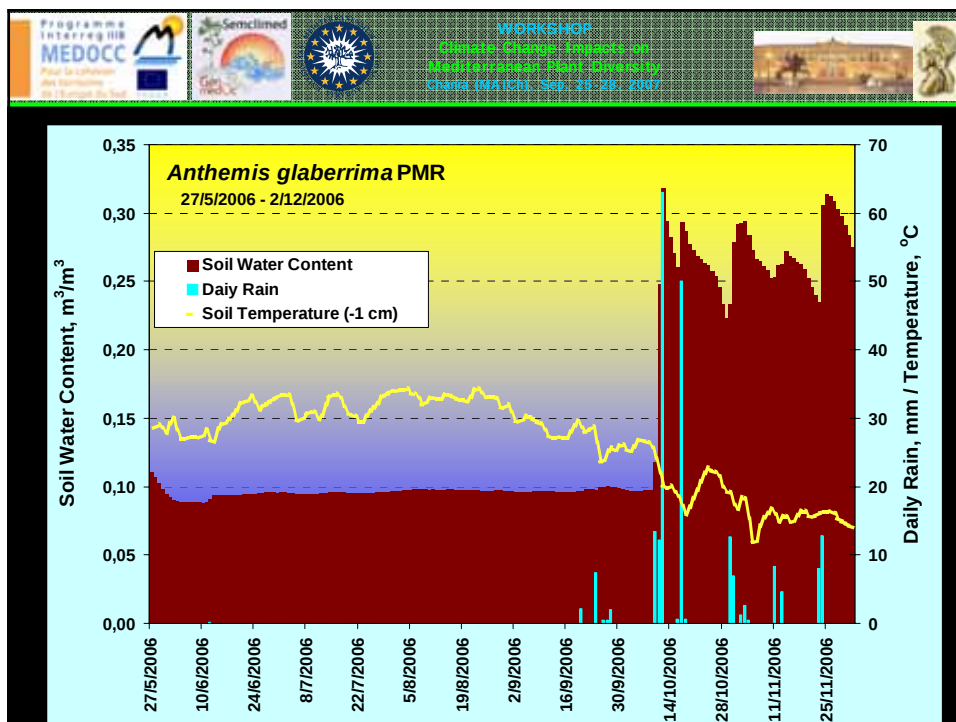
Anthemis glaberrima
(an endangered Cretan endemic annual)

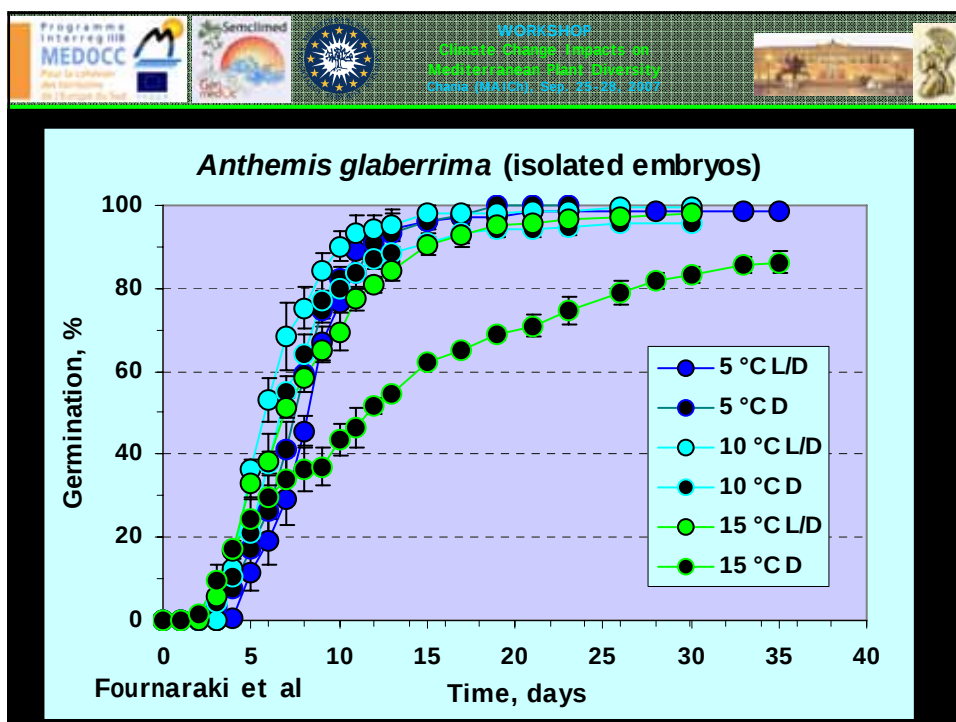
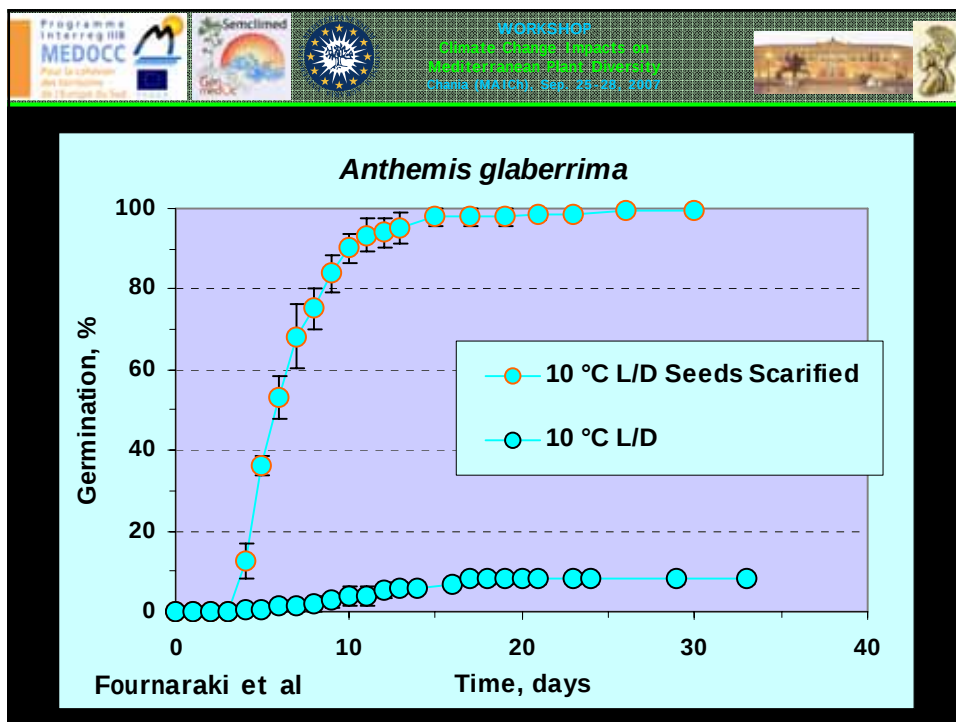


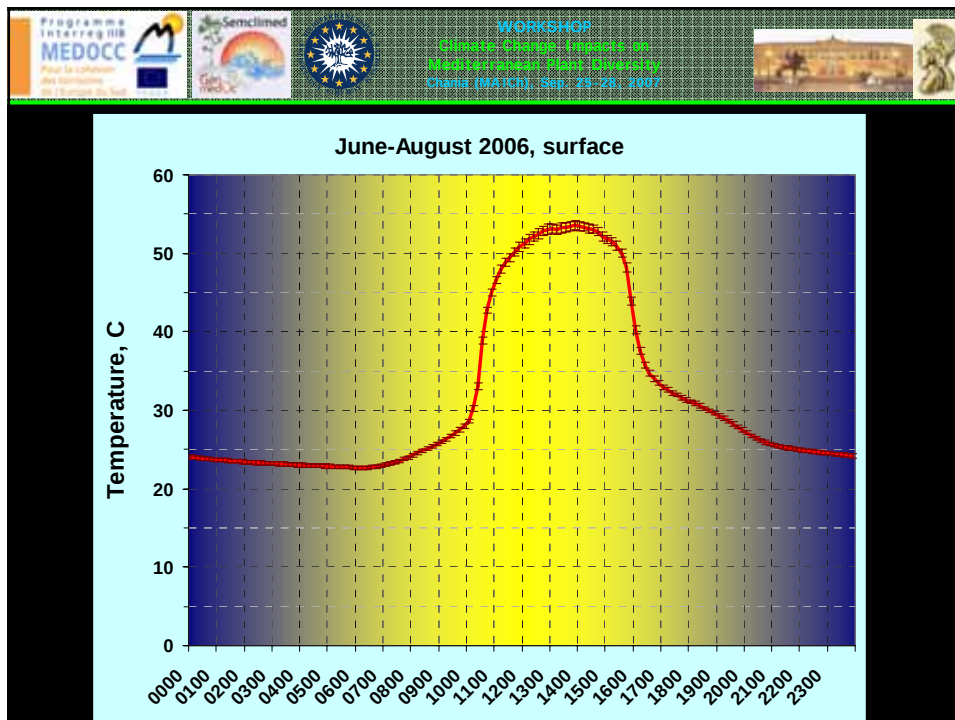












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WORKSHOP
ANALYSIS OF THE CLIMATE IN THE MEDITERRANEAN
CHANGING CLIMATE: Sep. 24-28, 2007

Anthemis glaberrima

UNANSWERED QUESTIONS

- ❖ ? The role of high, summer temperatures
- ❖ ? Presence of inhibitors in the seed coat
- ❖ ? A persistent soil seed bank

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Genetics, Ecology, Evolution
of the Mediterranean
Chania (Greece), Sep. 24-28, 2007

CASE STUDY V

Phoenix theophrasti
(Theophrastus' palm, a Cretan subendemic)

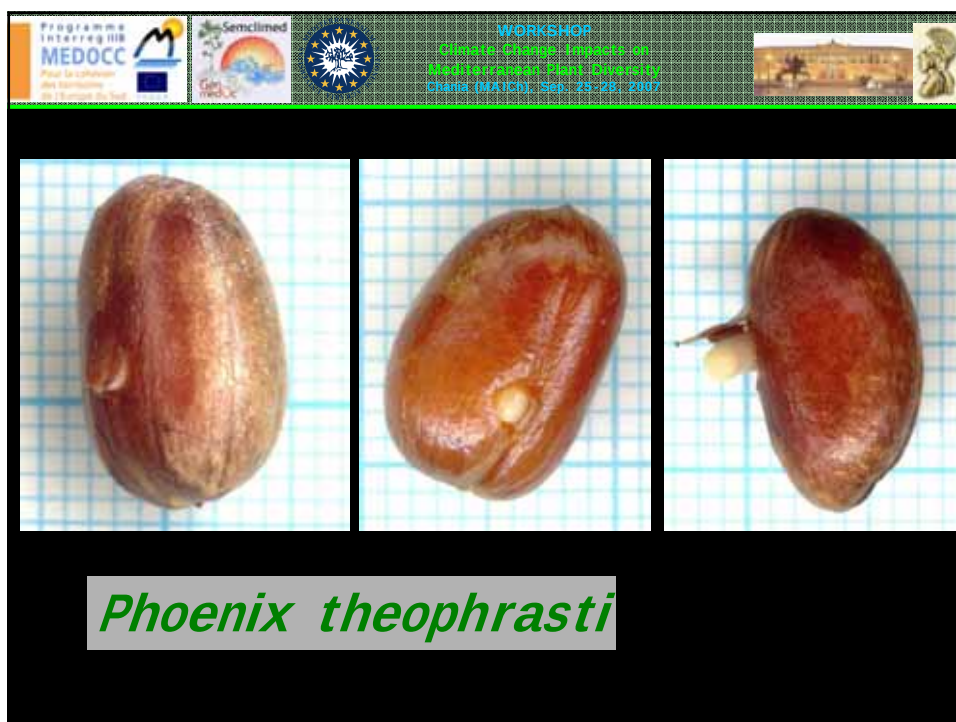
Programme Interreg IIIb MEDOCC
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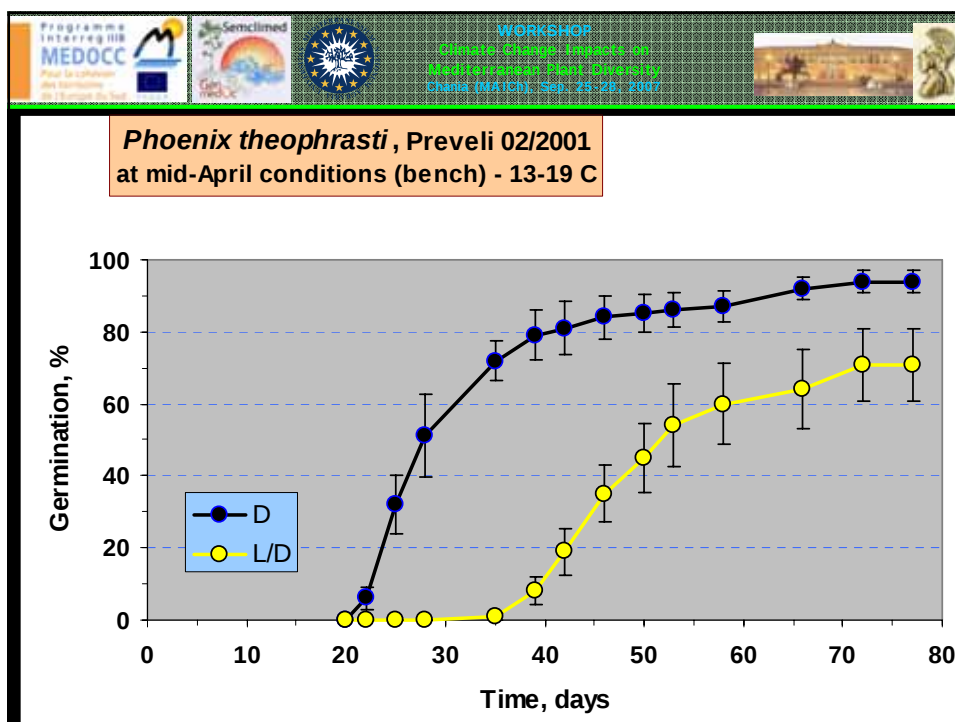
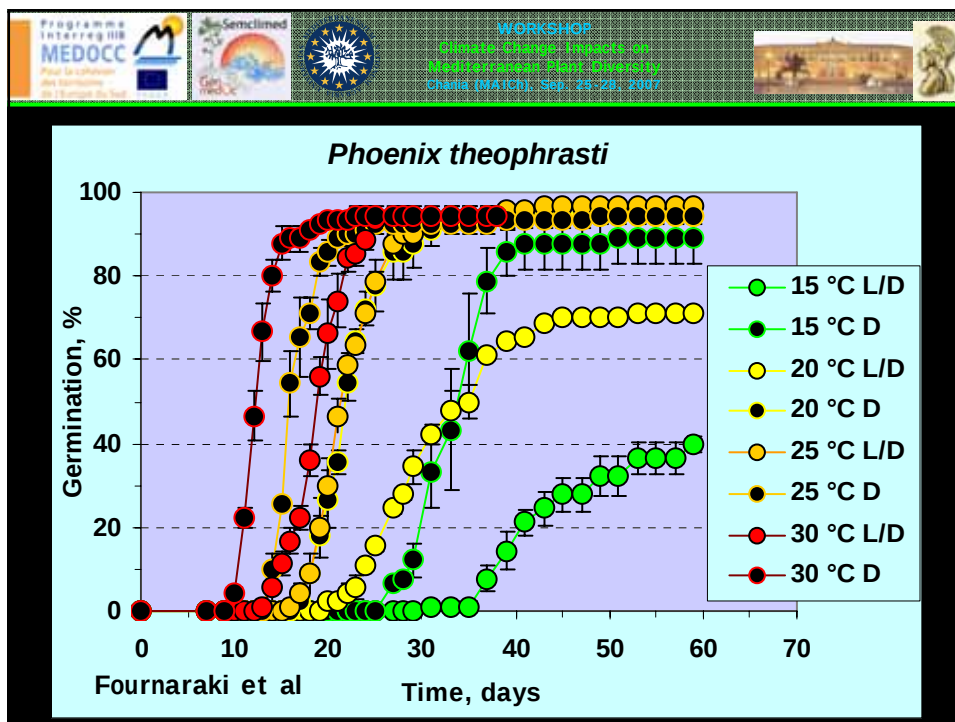
Phoenix theophrasti

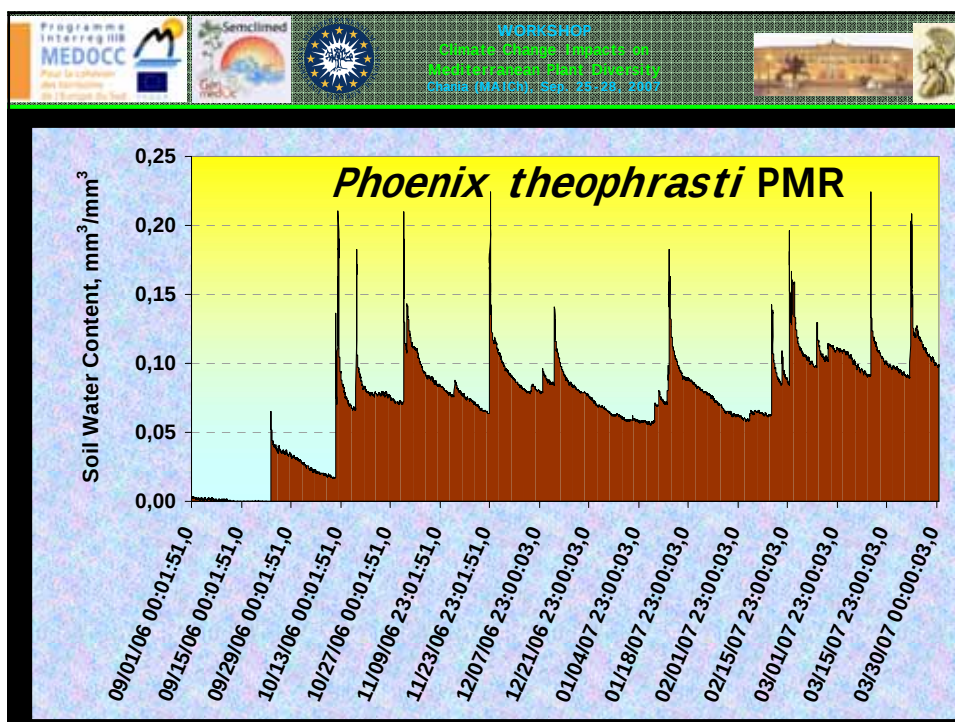
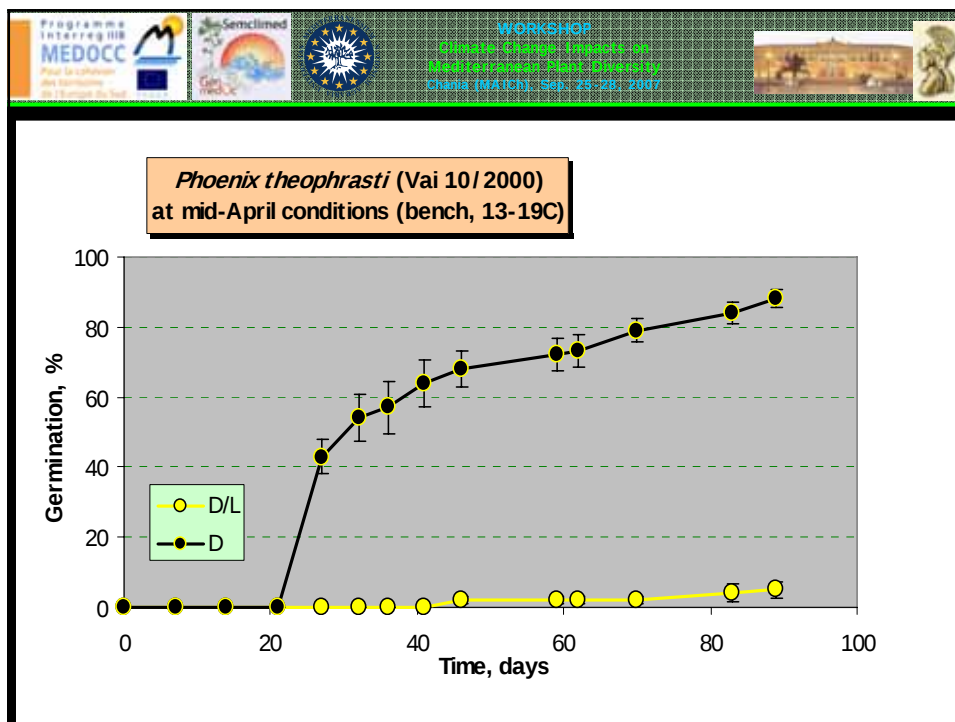


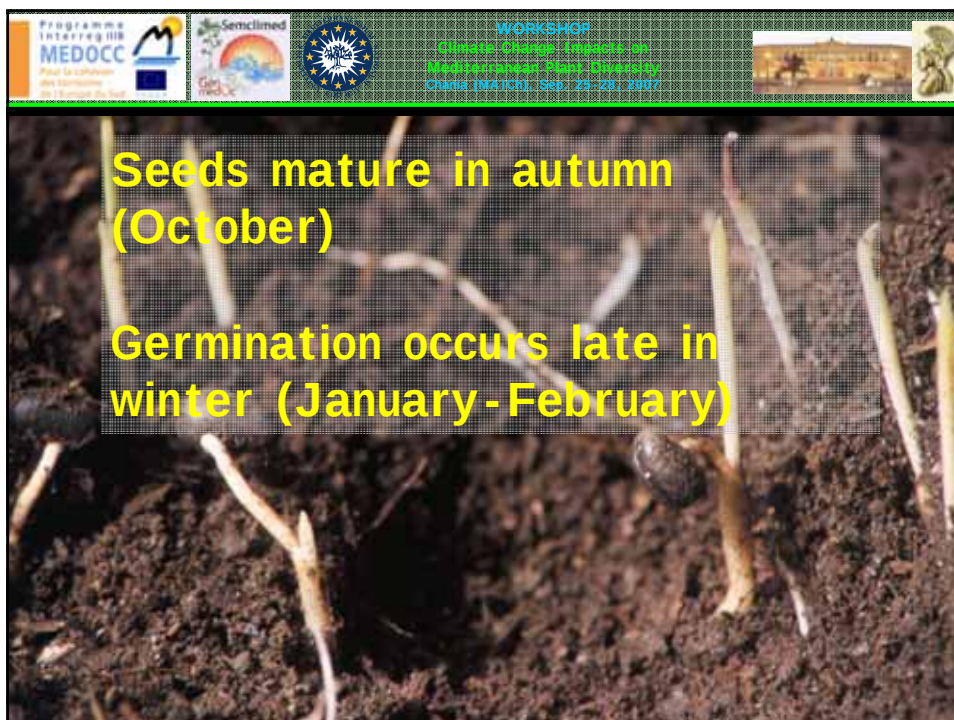
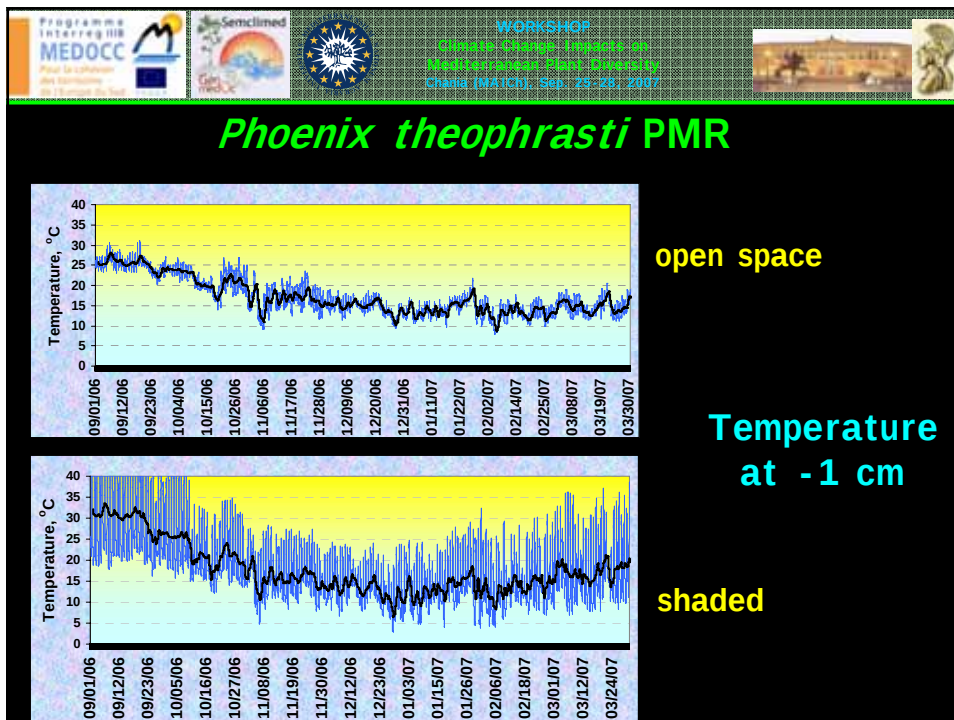























***Nepeta sphaciotica* P.H. Davis**

Une population seule,
 ca. 30000 individus
 à une surface
 moins de 4 ha

Critically Endangered: CR







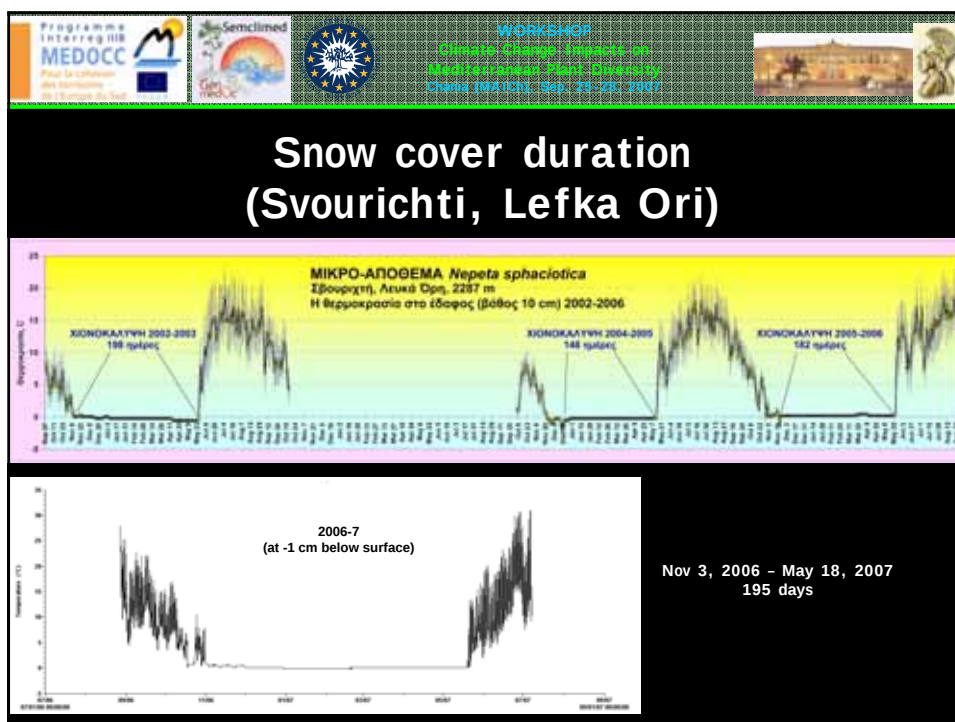
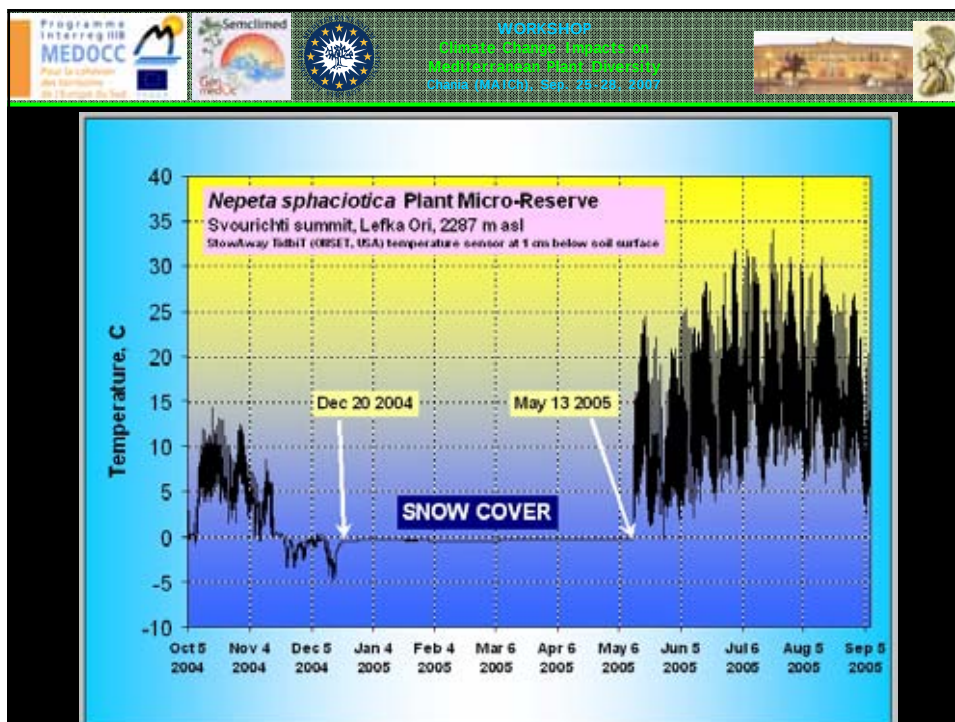


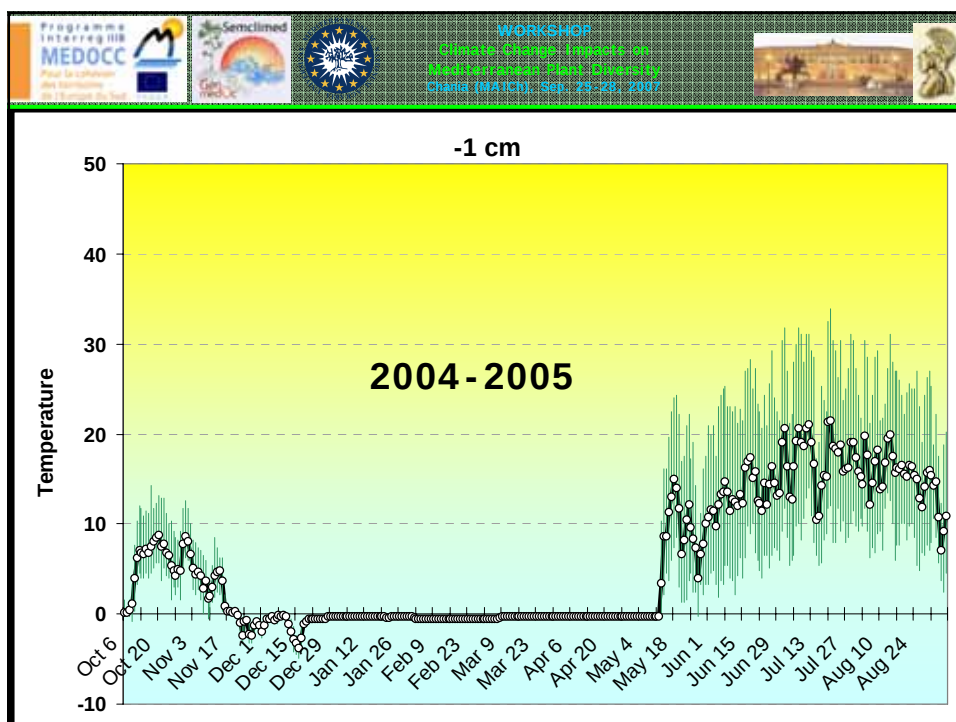
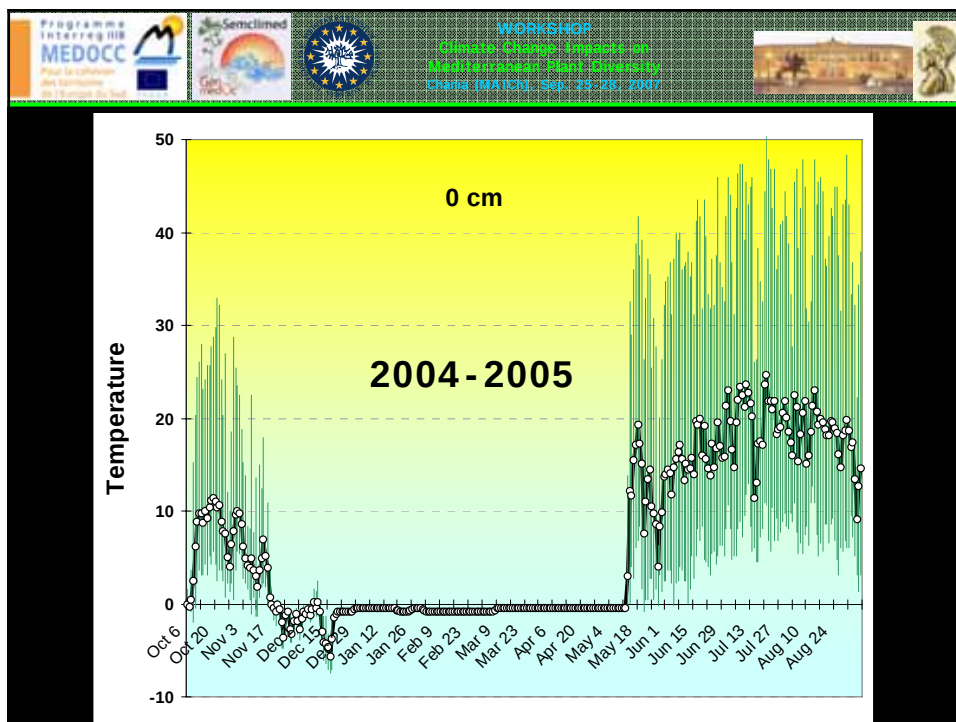

Nepeta sphaciotica

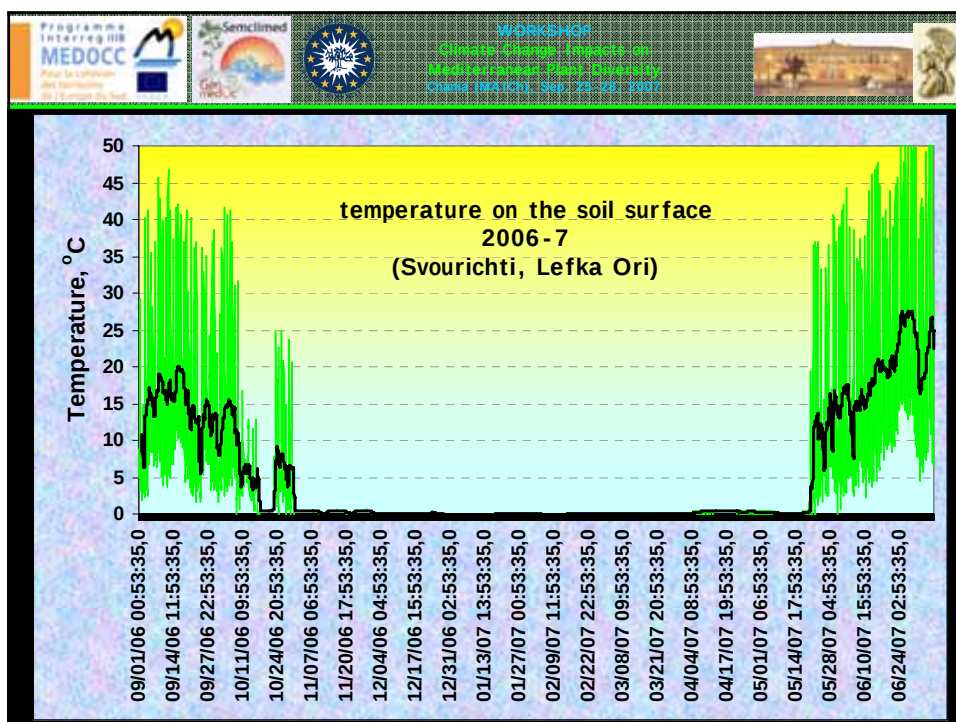
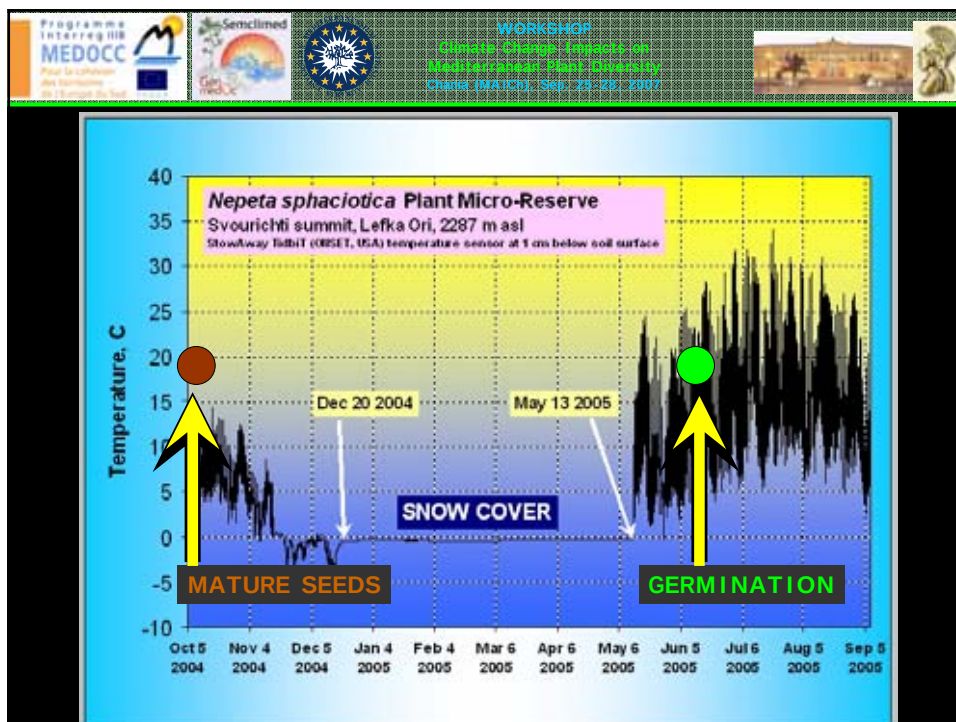


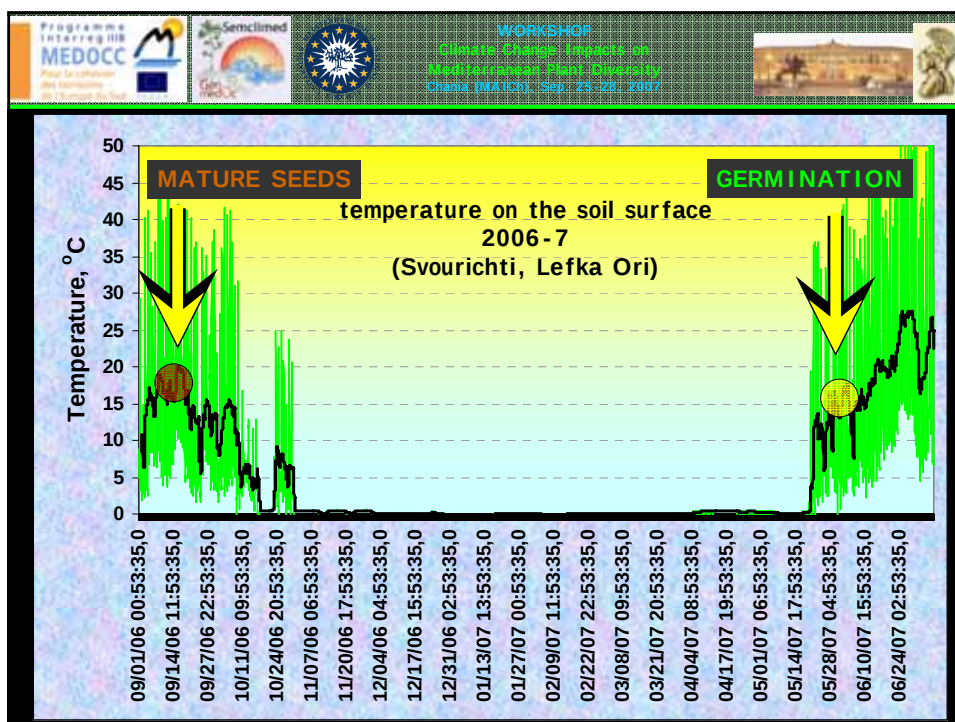
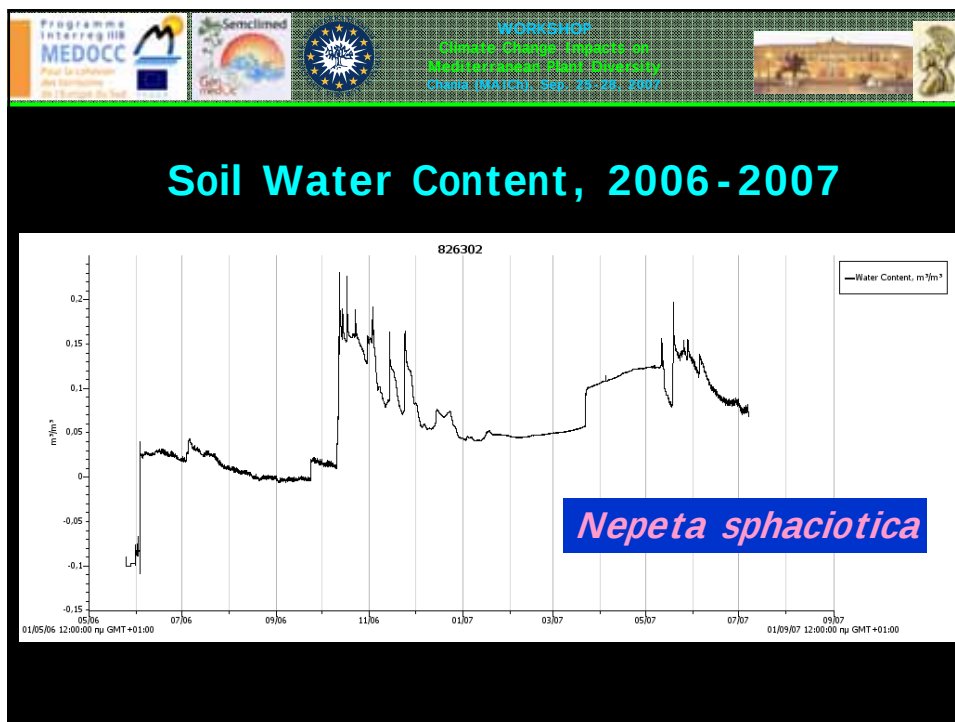


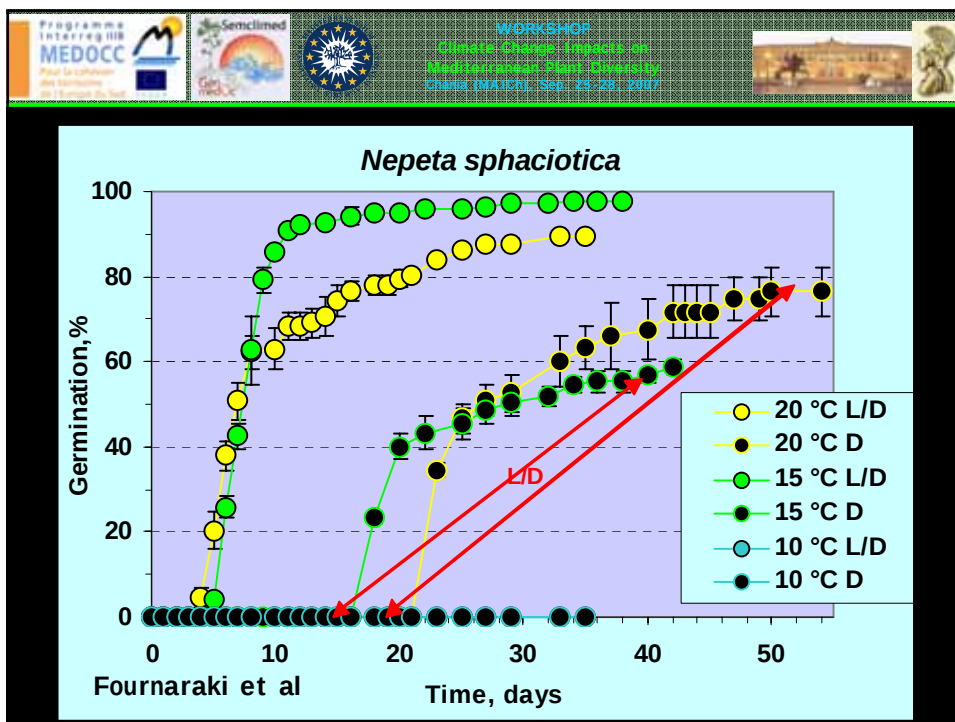
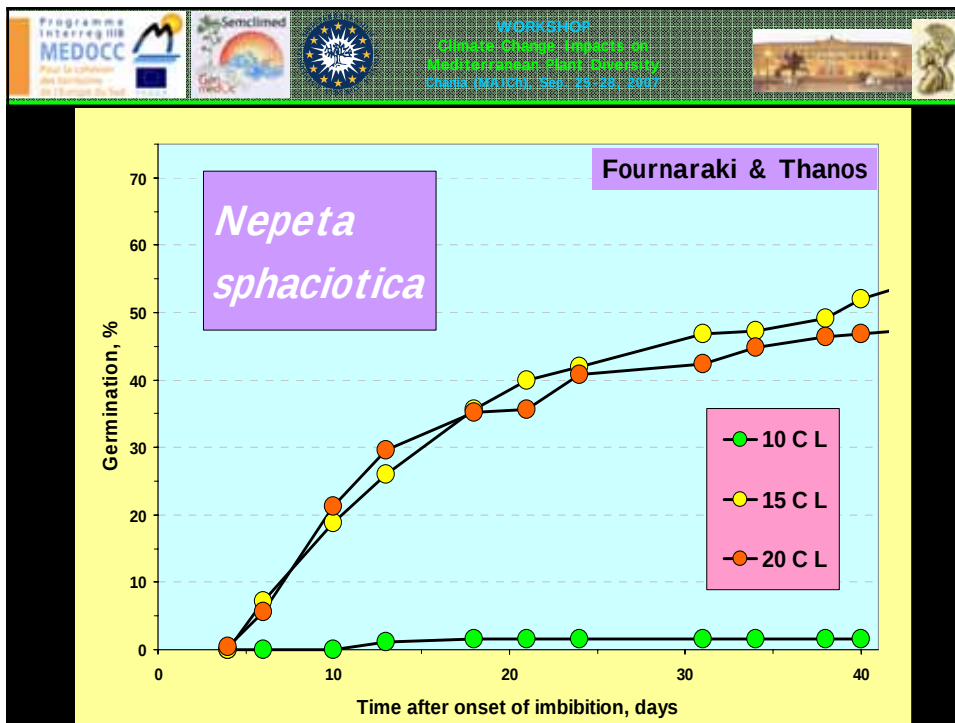


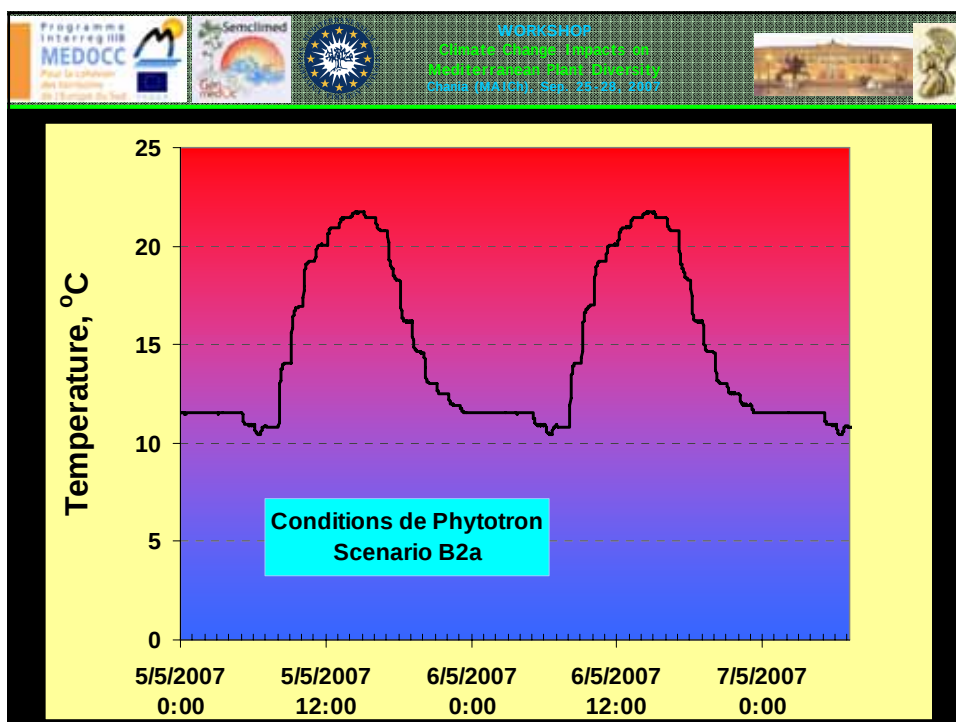
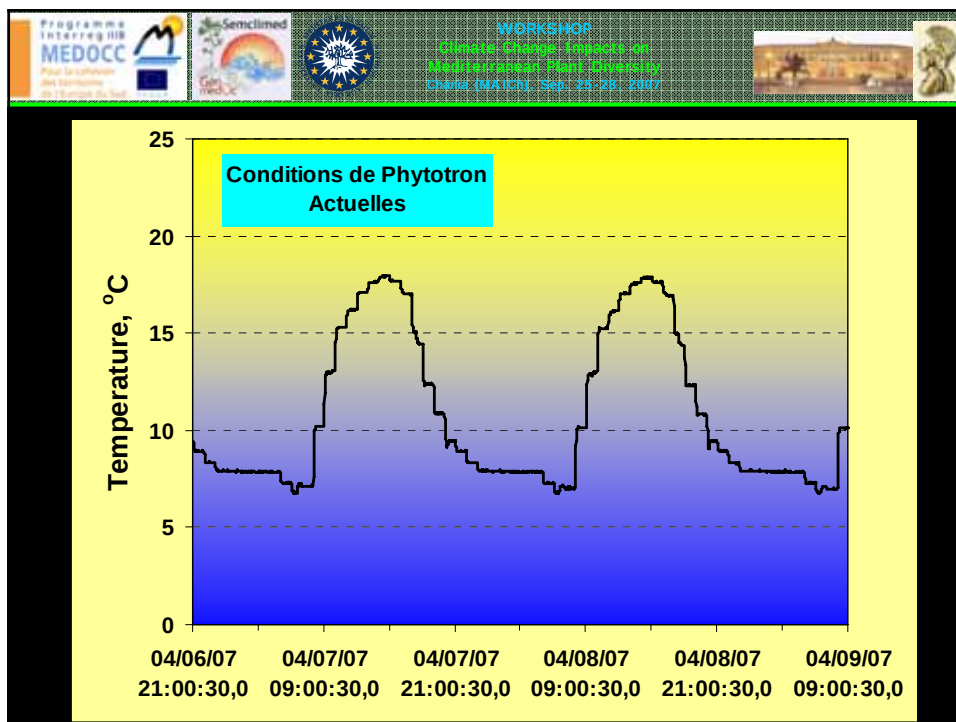


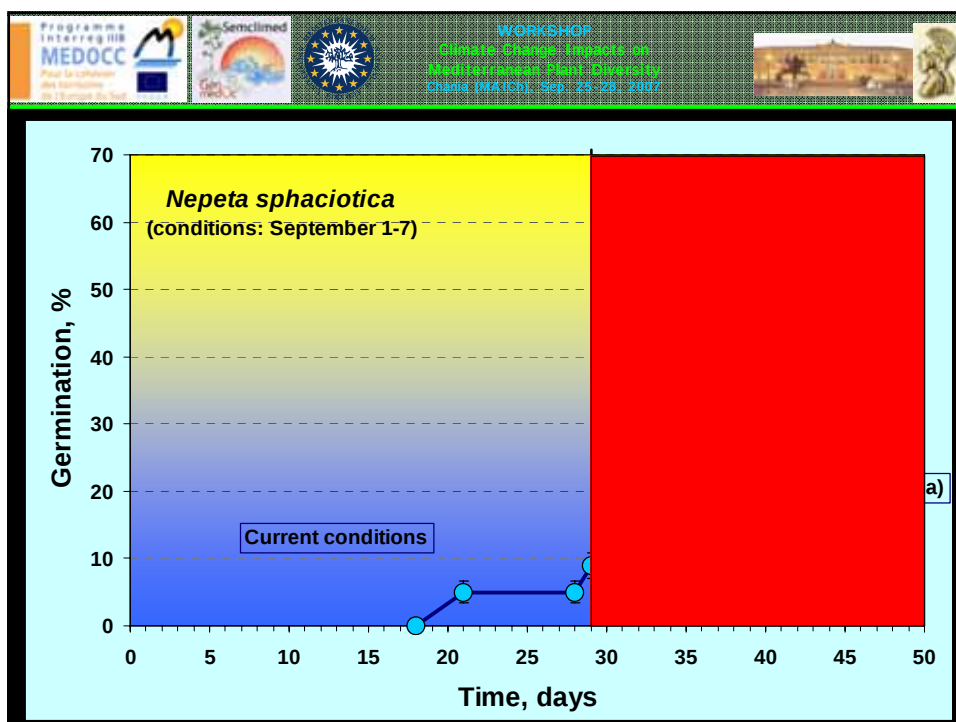
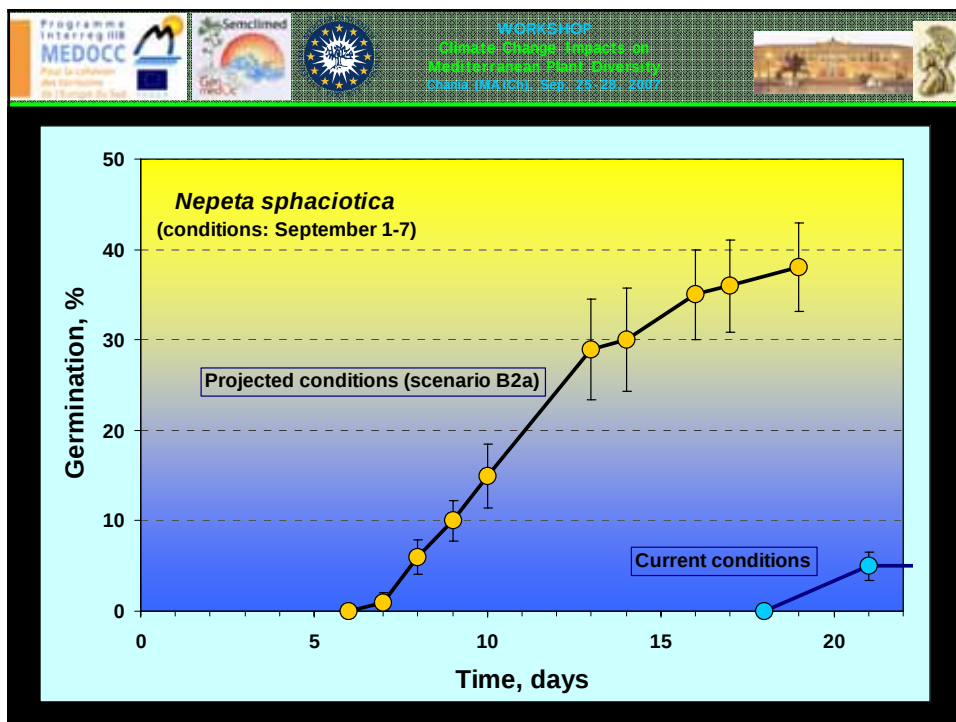


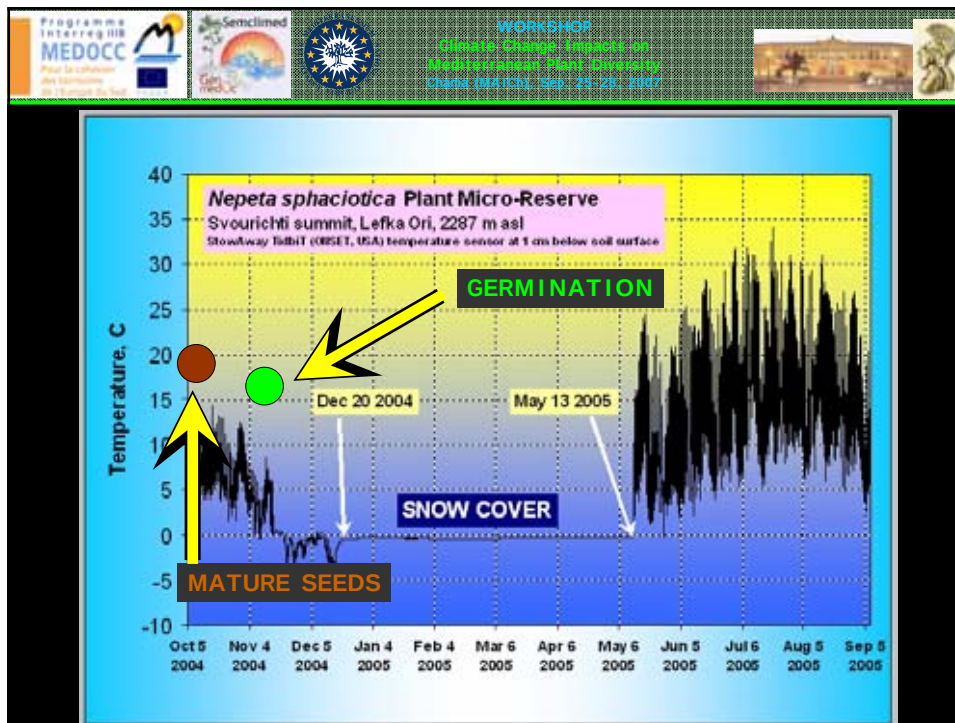












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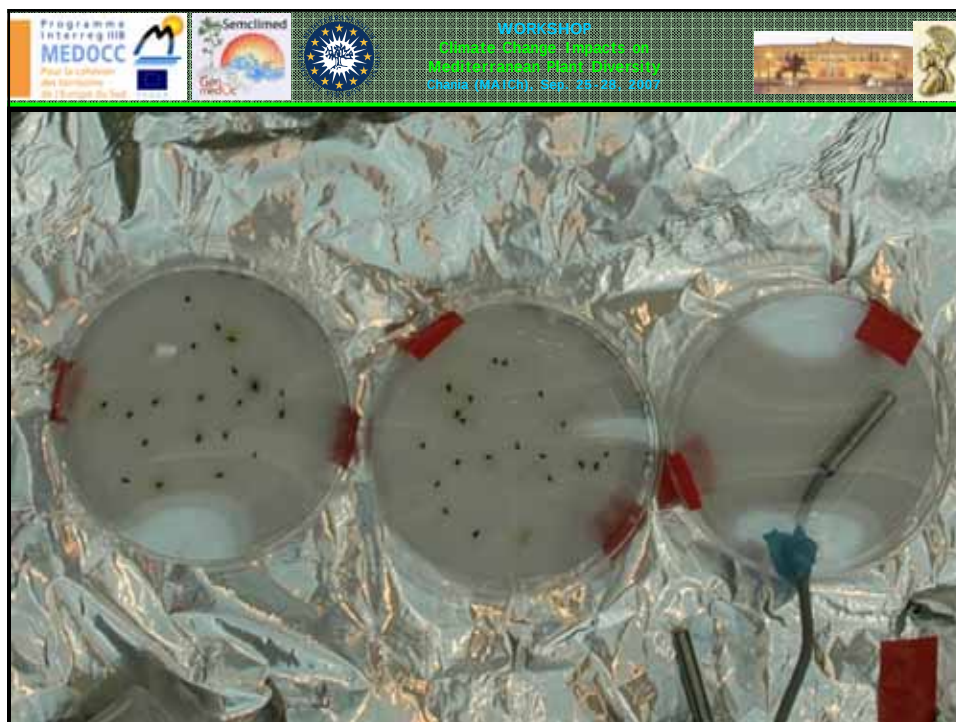
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MEDOCC Project: MEDOCC-04
MEDOCC Project: MEDOCC-04
Change (MATHON) Sep. 24-28, 2007

Technical and practical aspects







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Pour la cohésion des territoires
du Languedoc-Roussillon

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Chambre (MAG) - Sep. 24-28-2007

La température

La lumière

- photoperiode
- qualité
- quantité

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<http://aa.usno.navy.mil/aa/aa.html>
<http://aa.usno.navy.mil/aa/aa.html>
 China (WATC), Sep. 24-28, 2007

U.S. Naval Observatory Astronomical Applications Department

Table of Sunrise/Sunset, Moonrise/Moonset, or Twilight Times for an Entire Year

<http://aa.usno.navy.mil/>

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<http://aa.usno.navy.mil/aa/aa.html>
<http://aa.usno.navy.mil/aa/aa.html>
 China (WATC), Sep. 24-28, 2007

Form B - Locations Worldwide

<http://aa.usno.navy.mil/> (1 of 2) [11/5/2007 9:10:10 AM]

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Specify year, type of table, and place:

Year: Type of table:

Place Name:

The place name you enter above is used only in the table header; you can enter any identifier, or none (do not use punctuation characters).

Longitude: ☐ east ☒ west degrees minutes

Latitude: ☒ north ☐ south degrees minutes

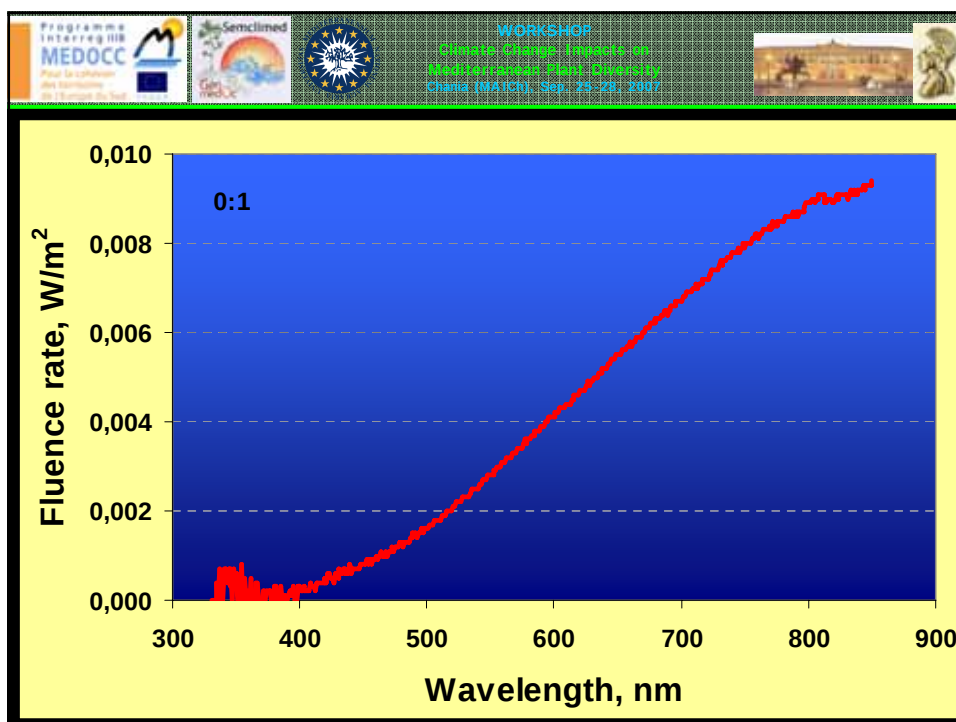
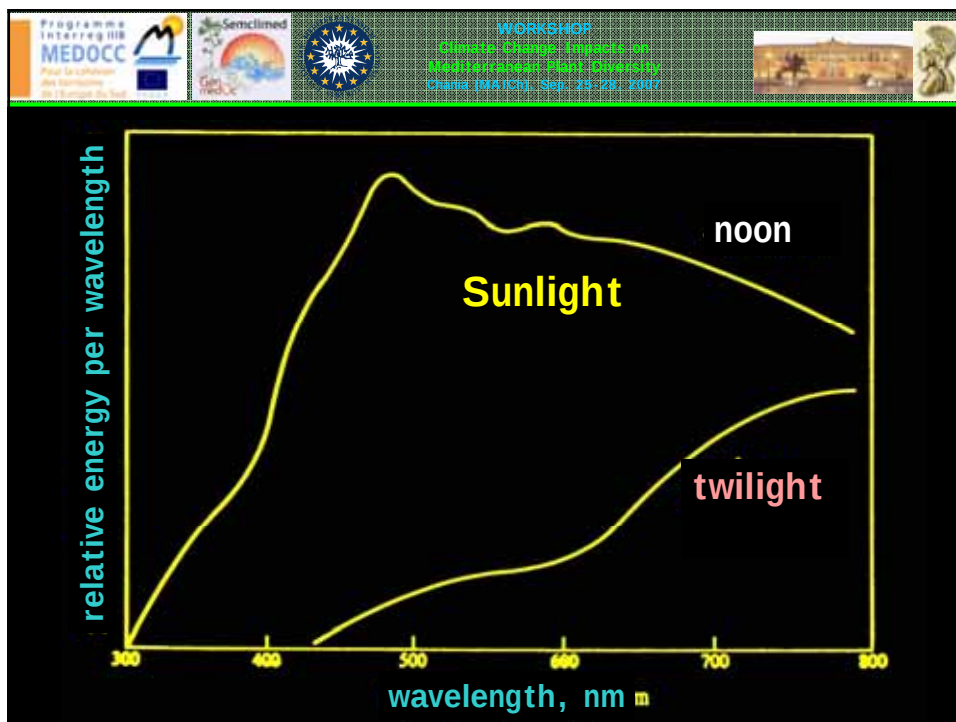
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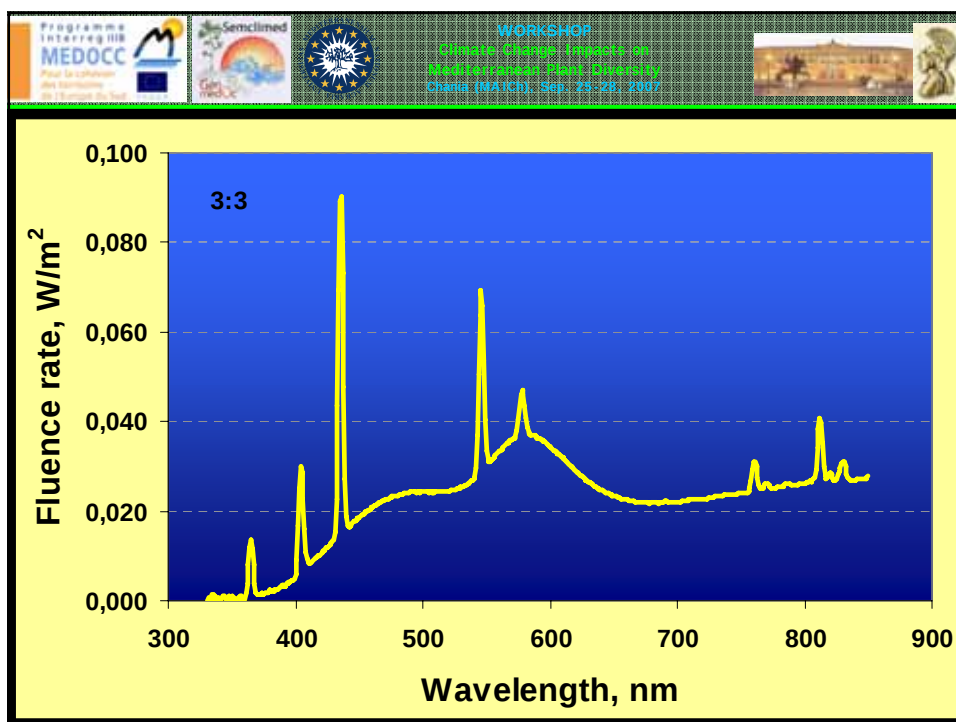
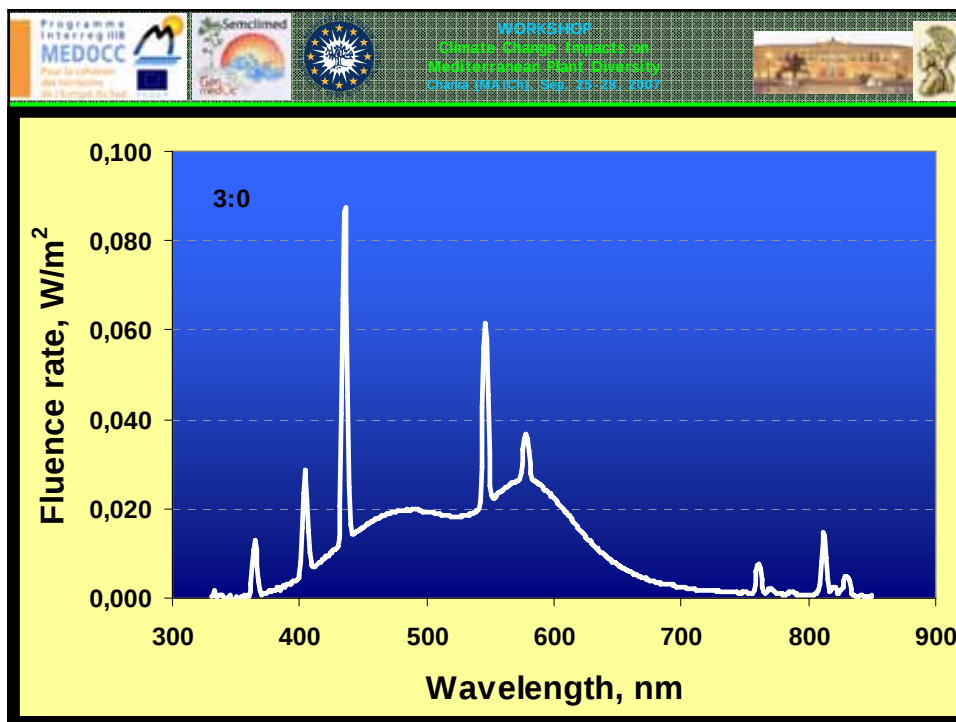
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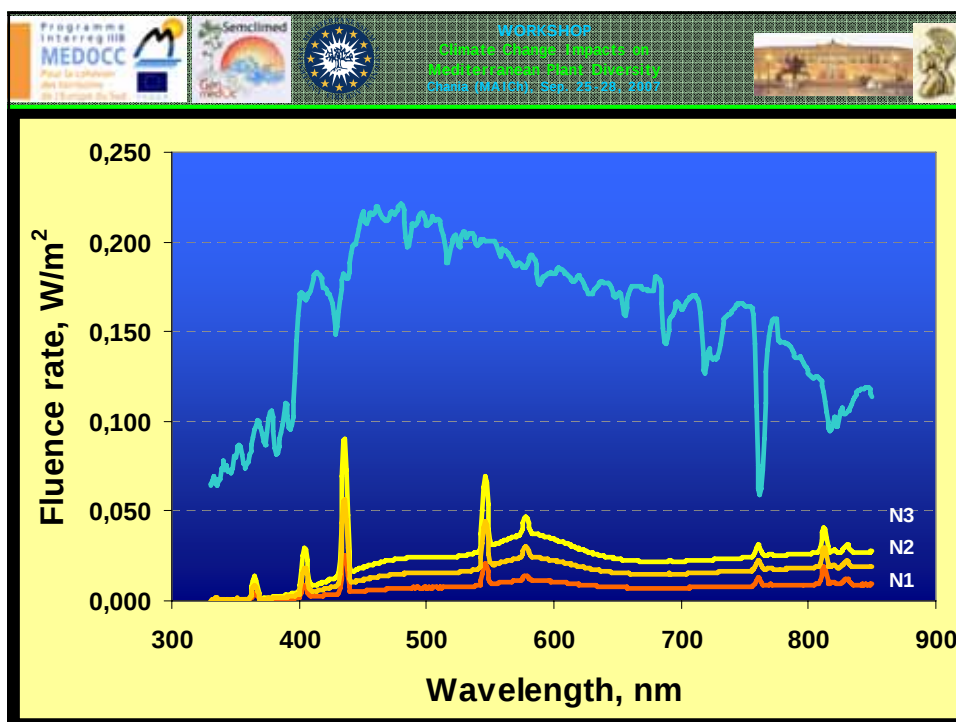
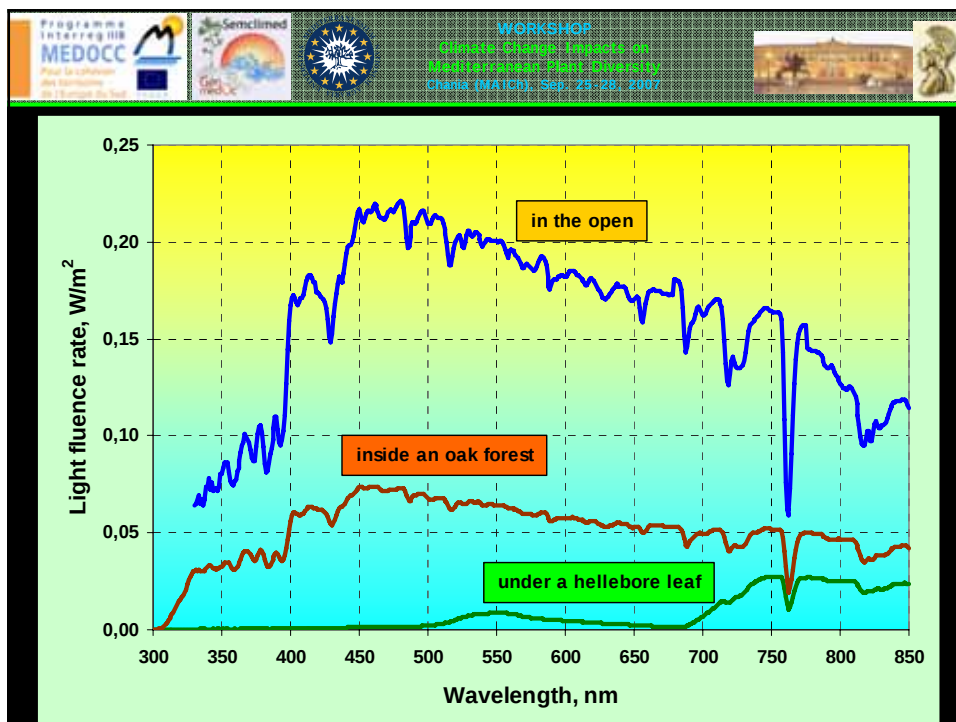
Need coordinates? Try NOAA's [GEOmet Harmonic Server](#) (click on the "GIS Search" link at the top of the left frame).

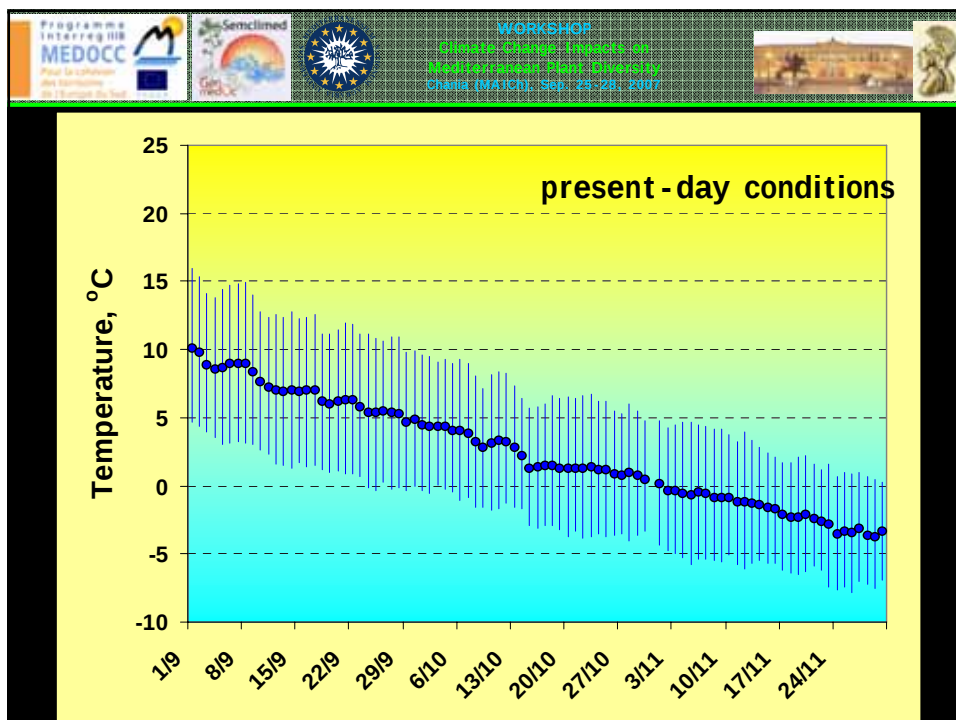
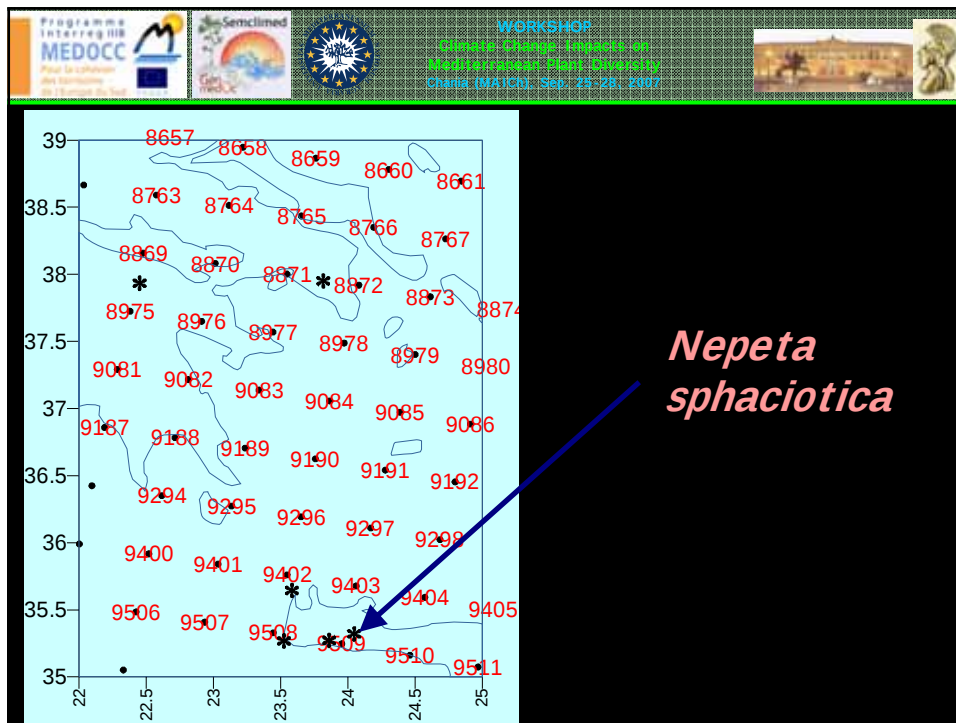
Need U.S. coordinates? Try this [USGS](#) web page.

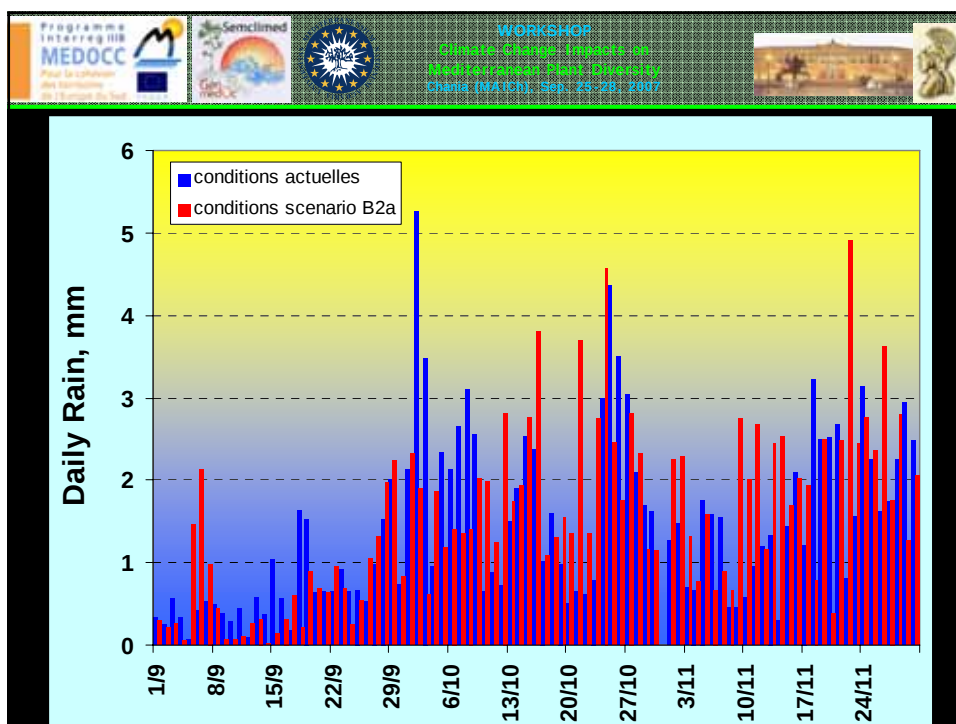
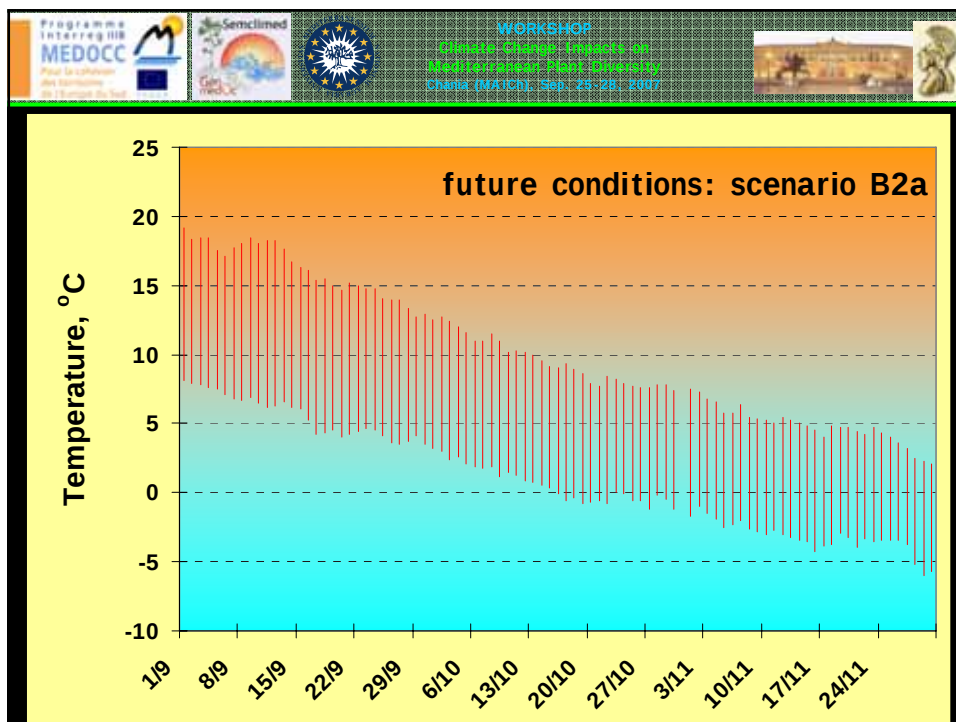
Need a time zone? Try the [time zone map](#).

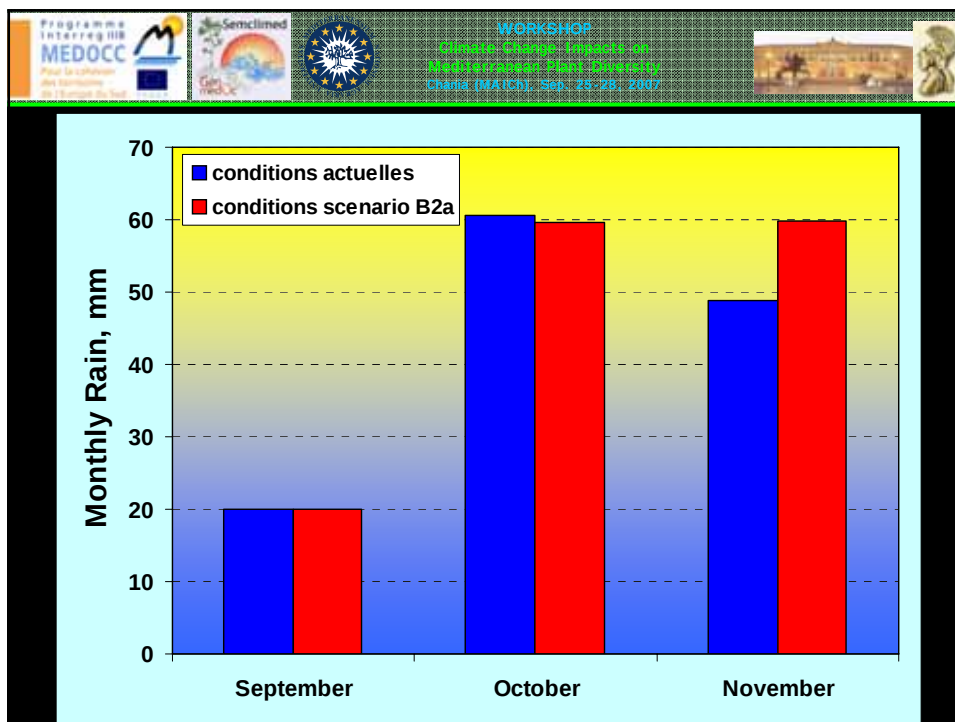












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du Sud







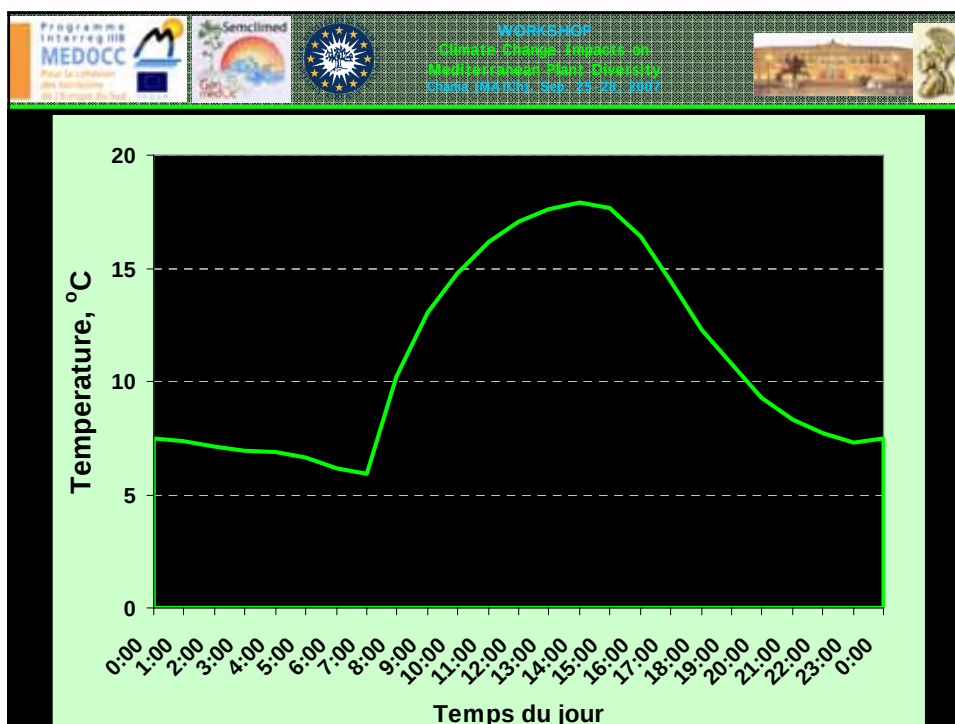
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Chania (MALTESE) - Sep. 24-28, 2007

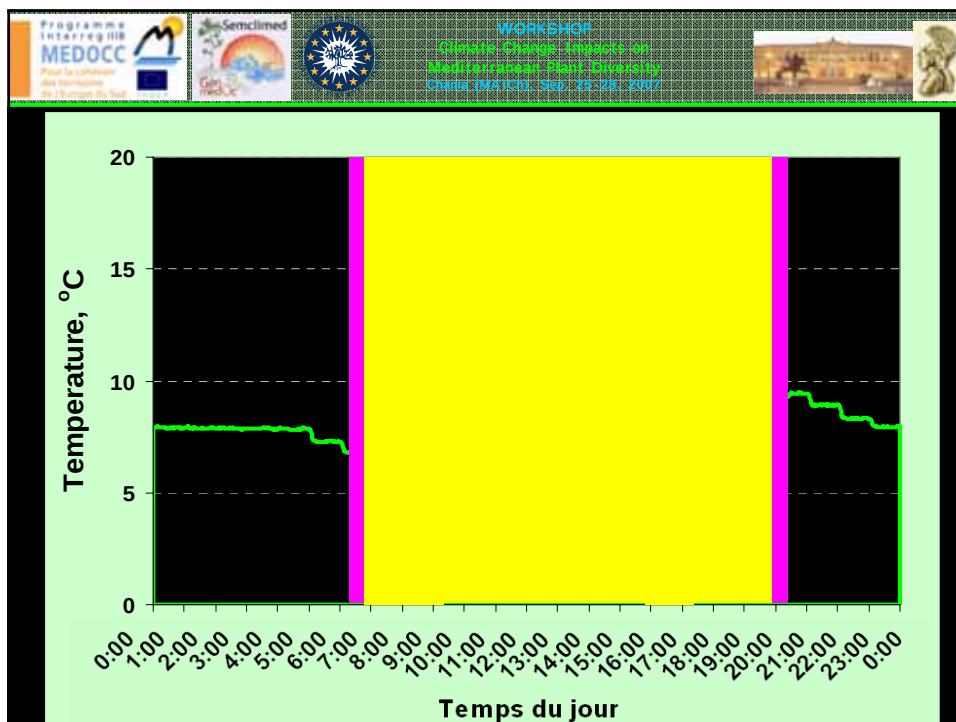
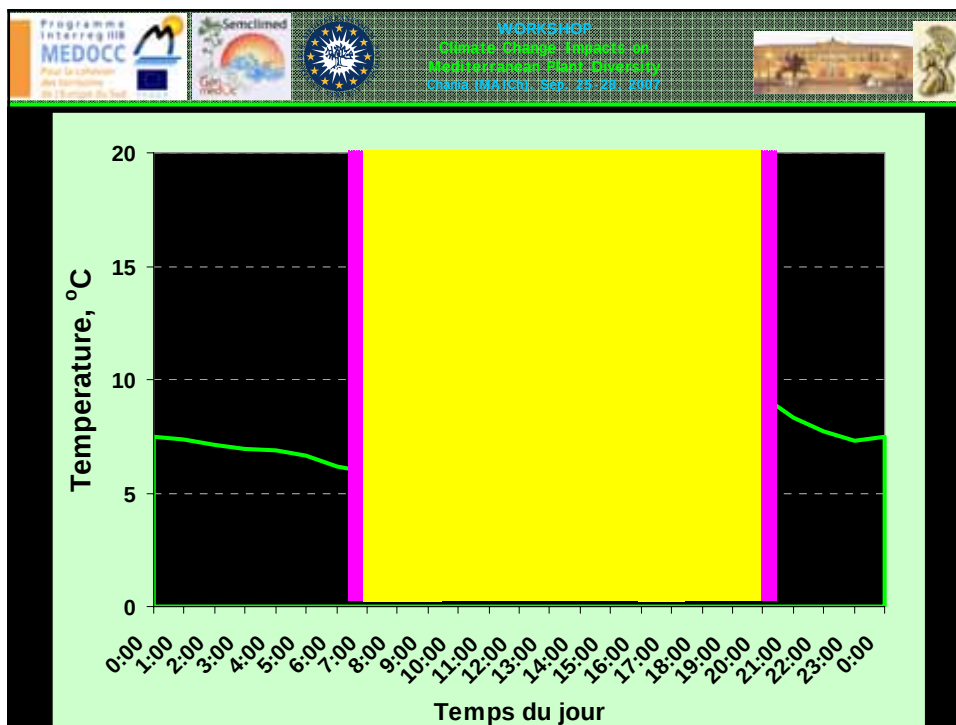


SVOURICHTI, 2280 m asl
PRESENT CONDITIONS (1-7 September)
at 1 cm below soil surface

	time	Temperature (controller)	Light		Light Temperature (in Petri dish)
			Fluorescent	Incandescent	
TWILIGHT SUNRISE	5:00	6,7	0	0	7,3
	6:00	6,2	0	0	6,8
	6:31	6,2	0	1	7,2
	6:57	6,0	1	1	7,1
	8:00	9,2	1	1	10,2
	9:00	12,0	1	1	13,0
	9:27	11,5	2	2	13,1
	10:00	13,8	2	2	15,3
	11:00	14,2	3	3	16,2
	12:00	15,1	3	3	17,1
SUNSET	13:00	15,6	3	3	17,6
	14:00	15,9	3	3	17,9
	15:00	15,6	3	3	17,6
	16:00	15,4	2	2	17,0
	17:00	13,4	2	2	15,0
	17:25	13,4	1	1	14,4
	18:00	11,3	1	1	12,4
	19:00	9,8	1	1	10,8
	19:48	9,8	0	1	10,8
	20:00	8,3	0	1	9,4
TWILIGHT	20:14	8,8	0	0	9,4
	21:00	8,3	0	0	8,9
	22:00	7,7	0	0	8,3
	23:00	7,3	0	0	7,9

Programme Interreg III MEDOCC Pour la cohésion des territoires du Sud-Est WORKSHOP Climate Change Scenario 2007-2020 Climate (M11) - Sep. 24-28, 2007					
SVOURICHTI, 2280 m asl B2a CLIMATE CHANGE SCENARIO (1-7 September) [+4 DEGREES] at 1 cm below soil surface					
	time	Temperature (controller)	Light		Light Temperature (in Petri dish)
			Fluorescent	Incandescent	
TWILIGHT	5:00	10,7	0	0	11,2
	6:00	10,2	0	0	10,7
SUNRISE	6:31	10,2	0	1	11,1
	6:57	10,0	1	1	10,9
	8:00	13,2	1	1	14,1
	9:00	16,0	1	1	16,8
	9:27	15,5	2	2	16,8
	10:00	17,8	2	2	19,0
	11:00	18,2	3	3	19,6
	12:00	19,1	3	3	20,6
	13:00	19,6	3	3	21,1
	14:00	19,9	3	3	21,5
	15:00	19,6	3	3	21,1
SUNSET	16:00	19,4	2	2	20,6
	17:00	17,4	2	2	18,6
	17:25	17,4	1	1	18,3
	18:00	15,3	1	1	16,2
	19:00	13,8	1	1	14,6
TWILIGHT	19:48	13,8	0	1	14,6
	20:00	12,3	0	1	13,3
TWILIGHT	20:14	12,8	0	0	13,3
	21:00	12,3	0	0	12,8
	22:00	11,7	0	0	12,2
	23:00	11,3	0	0	11,7









WORKSHOP

SEEDS: Ecology, Dispersal and Germination

Chania (Greece), Sep. 24-28, 2007




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RESEARCH OPINION

Mistakes in germination ecology and how to avoid them

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WORKSHOP

SEEDS: Ecology, Dispersal and Germination

Chania (Greece), Sep. 24-28, 2007




1. What should I try to germinate?

As far as possible, you should use the intact natural dispersal unit, because removal of parts from it may increase/decrease germination percentages. The 'intact natural dispersal unit' means the fruit or seed in the form in which it would normally occur in nature after dispersal. For fleshy fruits, this means removing the flesh, which often contains germination inhibitors. Unless it is likely to happen in the field, you should not attempt to remove structures such as the lemma and palea that remain tightly bound to many grass grains, even if their removal promotes germination.





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2. What conditions should I use?

It is best to test seeds/fruits over a range of alternating temperatures regimes, simulating those in the natural habitat during the growing season, and also place seeds/fruits in light and in darkness. If possible, constant temperatures and continuous light should be avoided; obviously, these conditions are not encountered frequently in nature, and in some species may decrease germination percentages. Also, high photon irradiance can inhibit germination. To answer this question, consider the sequence of environmental factors to which the seeds are exposed between maturity and germination. For example, if a seed is dormant at maturity in spring and germinates in autumn, it obviously does not need to be cold stratified.





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3. Why should fresh seeds be used when determining if seeds are dormant?

If seeds have non-deep physiological dormancy, after-ripening (dormancy break) can occur during dry storage, especially at room temperatures. In a few species, drying can cause seeds to enter dormancy. If seeds have physical dormancy, drying may cause the water gap to open. The key point is that if your seeds have already received any 'treatment' (however inadvertent) before you try to germinate them, you cannot know what the effects of this treatment have been.

5. When should a germination test be terminated?

Depending on the species, considerable dormancy break can occur in dry seeds at room temperatures within a few weeks; thus, we suggest that a test should be stopped within about 4 weeks. If you want to know if dormancy break and germination would eventually occur at a certain temperature regime, seeds could be checked at 2-week intervals and kept for years. Seeds at all test conditions should be given the same period of time for germination.

6. If fresh seeds germinate to a high percentage at only a portion of the test temperatures, are the seeds non-dormant?

Some freshly matured seeds are non-dormant, but they germinate only at high or at low temperatures, depending on the species. If the range of temperatures over which seeds will germinate cannot be increased, then seeds are non-dormant. However, in many species, the temperature range for germination will increase if seeds are given a dormancy-breaking treatment, e.g. cold stratification or exposure to high summer temperatures. If the temperature range increases, then the fresh seeds were conditionally dormant. The only way to know if fresh seeds are non-dormant (with a high or a low temperature requirement for germination) or conditionally dormant is to test them over a range of temperatures, give them a dormancy-breaking treatment and then test over the same range of temperatures. If the temperature range for germination increases, the fresh seeds were conditionally dormant. Try to make sure your test temperatures cover the whole range over which the seeds will germinate; even if your seeds germinate well at 25°C, test them at 30°C and 35°C before concluding that 25°C is the optimum temperature for germination.





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7. How can I tell if seeds have physical dormancy?

Physical dormancy means the seed (or fruit) coat is water impermeable. To determine if diaspores are water impermeable, they should be weighed before and after a period of incubation (e.g. 24h) on a wet substrate. Physical dormancy is currently known in the *Anacardiaceae*, *Bixaceae*, *Cannaceae*, *Cistaceae*, *Cochlospermaceae*, *Convolvulaceae* (including *Cuscutaceae*), *Curcubitaceae*, *Dipterocarpaceae* (subfamilies *Monotoideae* and *Pakaraimoideae*, but not *Dipterocarpoideae*), *Fabaceae* (all three subfamilies), *Geraniaceae*, *Malvaceae* (including *Bombacaceae*, *Sterculiaceae* and *Tiliaceae*), *Nelumbonaceae*, *Rhamnaceae*, *Sarcolaenaceae*, *Sapindaceae* and *Surianaceae*. If your species is not in one of these families, it probably has water-permeable seeds/fruits. It should be noted that not all members of all of these families have water-impermeable seeds. For example, in *Anacardiaceae* only members of the 'Rhus complex' have physical dormancy, and seeds of the other 70 or so genera in the family have either physiological dormancy or are non-dormant.





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10. If seeds are water permeable, why would mechanical or acid scarification promote germination?

Acid and mechanical scarification reduce the mechanical restriction of the seed/fruit coat. Thus, the radicle is able to emerge from the seed, although growth potential of the embryo has not been increased. Scarification also may overcome a light requirement for germination, reduce sensitivity to far-red light and increase sensitivity to gibberellic acid.





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12. How can I tell if seeds have morphological dormancy (MD)?

Seeds will imbibe water. They have a relatively small (low embryo length/seed length ratio) embryo in a relatively large seed with much endosperm. In MD, the embryo has to grow (inside the seed) before the seed germinates, and seeds germinate in about 4 weeks from the start of incubation. However, not every seed with a relatively small embryo has MD. To verify that seeds have MD, you need to demonstrate that the embryo grows inside the seed before the radicle emerges. To do this, measure the embryo in at least 15 freshly matured, imbibed seeds, and then measure the embryo in other seeds when the seed coat splits, i.e. just prior to radicle emergence.





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14. Cold stratification is reported to break dormancy in my seeds. Thus, I put dry seeds in a paper bag in the refrigerator at 5°C for 12 weeks, but none of them germinated, regardless of the test temperature. Why?

The seeds did not germinate because they were still dormant. Seeds that require cold stratification must be imbibed for dormancy break to occur. In addition, it is not a wise general practice to store unimbibed seeds in paper bags at any temperature; doing so allows the seed moisture to change continually as a function of atmospheric relative humidity (RH). Any physiological change that is moisture content dependent (such as after-ripening) will be subject to the RH variability.





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SEEDS: FROM DARKNESS TO
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17. In my experiment, dark-incubated seeds were exposed to dim white (or room) light for about 2–3 min each day while germination was being checked. My results show that germination percentages were equally high in both light and darkness; thus, can I conclude that the seeds do not have a light requirement for germination?

No. During the brief period of exposure to light, the light requirement for germination may have been fulfilled in the 'dark-incubated' seeds. Only a few seconds of light are required to fulfil the light requirement of seeds of some species.





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SEEDS: FROM DARKNESS TO
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18. How safe are green safe lights for checking germination of dark-incubated seeds?

It depends on the species, time of year (i.e. stage of after-ripening) and quality of the light. Green light actually promotes seeds of some species to germinate, while seeds of other species are more sensitive to green light at some times of the year than at other times. The quality of the green light is important. It is not easy to get a narrow-band green safe light. Thus, at different stages of dormancy break, it is important to compare germination of seeds exposed to the green light with that of seeds not receiving any green light. A green safe light may not stimulate germination in the early stages of dormancy break, but may stimulate germination in the late stages of dormancy break, i.e. when the seeds are more sensitive to germination-stimulating factors.





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**Green Seeds: Ecology and
 Conservation**
 CHANIA (MALTA), Sep. 24-28, 2007




19. Is it OK to collect the seeds, although they still look a little green?

Testing green seeds may not give you a true picture about seed dormancy, but you might get seedlings (if that is your aim), depending on the species. One way to avoid the problem of physical dormancy in seeds of some species is to collect seeds/fruits before they dry on the mother plant, i.e. before the seed/fruit coat becomes water impermeable during the drying process. In a few species whose seeds have physiological dormancy, picking green seeds can also be a way of avoiding the problem of dormancy. However, some seeds with physiological dormancy may never germinate if they are not allowed to dry. Sometimes, seeds can be picked while slightly immature, and they will be normal with respect to dormancy. If seeds must be collected before they are fully ripe, it is best to leave the seeds attached to as much plant material as possible and to store them in a damp place where they will dry slowly.





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21. If leaching in running water increases germination percentages, does this mean a chemical inhibitor was removed from the seeds?

Maybe, but maybe not. Proving that a particular compound is present and that it prevents germination *in situ* is very difficult. Further, depending on the temperature and on length of the leaching period, seeds may have actually received a period of warm or cold stratification.

